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ErZhiTianGui Decoction alleviates age-related ovarian aging by regulating mitochondrial homeostasis and inhibiting ferroptosis

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Abstract

Aim This study was designed to investigate the pharmacological effects and mechanisms of ErZhiTianGui Decoction (EZTG) for age-related ovarian aging in mice.

Methods This study used naturally aging mice as a model, and EZTG was used for intragastric administration. Ovarian pathological changes, as well as follicular reserve were assessed by hematoxylin and eosin staining, and serum hormone levels (anti-mullerian hormone, follicle-stimulating hormone), mitochondrial reactive oxygen species (ROS) and mitochondrial DNA (mtDNA) damage marker 8-hydroxy-2'-deoxyguanosine(8-OHdG), and lipid peroxidation markers glutathione(GSH) and malondialdehyde(MDA) were determined by enzyme-linked immunosorbent assay. Mitochondrial membrane potential (MMP) levels in ovaries were determined using flow cytometry. The levels of PINK1 and Parkin were observed using immunofluorescence staining. Mitochondrial-derived vesicles (MDVs) and mitochondrial morphology were observed using electron microscopy. Prussian blue staining was used to observe iron ion aggregation in ovarian tissue. The Iron assay kits detected total iron levels. Western blot was used to detect the expression of proteins related to mitochondrial and ferroptosis related genes.

Results After EZTG treatment, aged mice showed increased ovarian reserve, improved serum hormone levels, increased MMP, GSH levels, and decreased mitochondrial ROS, 8-OHdG, and MDA levels. Immunofluorescence staining showed decreased levels of PINK1 and Parkin, and the same trend was observed for the Western blot. Meanwhile, electron microscopy showed that EZTG improved the mitochondrial morphology in the ovaries of aged mice. EZTG also decreased the total iron and protein levels of Acyl-CoA synthetase long-chain family4 (ACSL4) and increased the protein level of glutathione peroxidase 4 (GPX4) in the ovaries of aged mice.

Conclusions EZTG can maintain PINK1/Parkin-mediated mitochondrial homeostasis, reduce the lipid peroxidation caused by the accumulation of ROS, and inhibit the occurrence of ferroptosis and delaying ovarian aging. These findings suggest that EZTG may be a promising drug for treating age-related ovarian aging in females.

Keywords ErZhiTianGui Decoction, Ovarian aging, Mitochondrion, Mitophagy, Ferroptosis

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Introduction

Age alone has an effect on fertility, women older than 35 are traditionally defined as women with advanced maternal age (AMA) [1], whose fertility declines significantly after the age of 35, with the most significant decrease in ovarian reserve. With the extension of life expectancy and the postponement of fertility intentions worldwide, the proportion of AMA has increased substantially [2, 3]. The mechanism of age-related ovarian dysfunction is not yet clear, and it is currently mainly related to factors such as mitochondrial dysfunction, free radicals and antioxidant systems, telomere and telomerase changes, and cell apoptosis [4]. Exploring the pathogenesis and potential treatment strategies of age-related ovarian aging is currently one of the hotspots in obstetrics and gynecology.

Mitochondria play a crucial role in ovarian aging and reproductive longevity. It has been clarified that mitochondrial dysfunction leads to ovarian aging [5]. Mitochondria are the main source of free radicals and ROS in cells and play a central role in oxidative phosphorylation and redox. The mitochondrial dysfunction produces excessive ROS, and a considerable accumulation of ROS leads to oxidative stress, which in turn affects mitochondrial homeostasis and forms a vicious cycle [6, 7]. Meanwhile, Mitochondria regulate various forms of cell death and play a crucial role in the regulation of ferroptosis [8, 9]. Ferroptosis, an iron-dependent form of non-apoptotic cell death in 2012, has seen exponential growth in research over the past few years [10]. Ferroptosis has also made some research progress in female reproduction, including its role in ovarian cancer [11], endometriosis [12], and polycystic ovary syndrome [13]. However, ferroptosis in age-related ovarian aging has not been elucidated. Regulation of mitochondrial homeostasis may alleviate the occurrence of ferroptosis, thereby alleviating ovarian aging.

Herbal medicine has been used for thousands of years to treat infertility, and they have shown good clinical effects in age-related ovarian aging [14]. Traditional Chinese medicine believes that the kidney is the root of reproduction, and deficiency of the kidney energy is the fundamental cause of age-related ovarian aging. EZTG Decoction, composed of ten herbal ingredients, have the effect of nourishing kidney-qi and tonifying kidney-yin [15]. EZTG has been used in clinical practice for over 20 years with good clinical effects in alleviating ovarian aging and improving pregnancy outcomes for elderly women [16]. To date, no study has been reported on the role of EZTG in alleviating ovarian aging. Therefore, this study used naturally aging mice as a model and administered EZTG intragastrically to explore the role of EZTG in alleviating ovarian aging.

Methods and material

Ethical statement

All animal experiments were conducted in strict accordance with the Guide to the Management and Use of Laboratory Animals issued by the National Institutes of Health. Animal Ethics Committee of Affiliated Hospital of Shandong University of Traditional Chinese Medicine approved all the experimental procedures used in this study (Approval number: 20,220,316).

Animal models and treatment

Sixty C57/BL6 female mice aged 36 weeks were obtained from JinanPengyue-laboratory animal breeding Co., Ltd. (SYXK20180015, Jinan, China). All mice were housed in standard conditions with a 12-h light/dark cycle at 22 ± 2 °C and a humidity of $55\pm 5\%$ with free access to food and water. We calculated the dose of EZTG to be taken by mice in the ratio of 9:1 According to the medication instructions, the therapeutic dose of EZTG for a woman weighing 60 kg is 18 g/day. The 60 mice aged 36 weeks were randomly distributed into three groups: an aged control group (old group), a low-dose EZTG group (EZTG-L, 2.7 g/kg/day by gavage), and a high-dose EZTG group (EZTG-H, 8.1 g/kg/day by gavage). The 20 mice aged 8 weeks were used as a young control group. The EZTG Decoction are produced by the Drug Manufacturing Unit of the Affiliated Hospital of Shandong University of Traditional Chinese Medicine, batch number 01-FZ032-03. The composition traditional Chinese herbs of EZTG Decoction include: *Cuscuta chinensis* (Tu Si Zi), *Ligustrum lucidum* (Nv Zhen Zi), *Herba Ecliptae* (Mo Han Lian), *Lycium chinense* Mill (Gou Qi Zi), *Angelica sinensis* (Dang Gui), *Radix Rehmanniae Preparata* (Shu Di Huang), *Ligusticum wallichii* (Chuan Xiong), *Paeonia lactiflora* Pall (Bai Shao), *Rhizoma cyperi* (Xiang Fu), *Radix Glycyrrhizae Preparata* (Zhi Gan Cao). The EZTG group was gavaged for 12 weeks, while the control group was gavaged with equivalent doses of normal saline. All mice were anesthetized and euthanized, and their blood and ovaries were collected for subsequent use, and the weight of mice and wet weight of ovaries were measured (ovarian index = ovarian mass/body weight, mg/g).

Histopathology

Ovaries were harvested and fixed with paraformaldehyde at a concentration of 4% for more than 24 h, and dehydration, wax leaching, and embedding were performed. Ovaries were cut into 4 μ m sections, which were dewaxed with ethanol and stained with Haematoxylin and eosin dyes. The number of Primordial follicles, Primary follicles, Secondary follicles, and Mature follicles is recorded based on morphological characteristics.

Serum hormone levels

Serum samples obtained before the mice were euthanized were centrifuged at room temperature for 20 min at 3000×g. Serum hormone levels, including anti-Müllerian hormone (AMH), and follicle-stimulating hormone (FSH), were measured using ELISA kits (F9402-A, F2555-A, Shanghai Fanke Industrial Co., Ltd, Shanghai, China), according to the manufacturer's instructions.

Mitochondrial membrane potential flow cytometric assay

Ovarian tissue was cut into small pieces, digesting it completely with trypsin, adding complete culture medium, and collecting the cell suspension. MMP levels were detected using the JC-1 assay kit (E-CK-A301, Elabscience). The cell suspension were removed in each group, and 1 mL JC-1 working solution was added after washing with PBS. Negative and positive control groups were set according to the instructions. The cell suspension stained with JC-1 staining buffer and detected by flow cytometry.

Mitochondrial ROS assay

ROS assay kits (AO036-100T/96S, Yishijiu Biotechnology Co., Ltd, Jiangsu, China) were utilised to quantify ROS levels in ovary tissue, according to the manufacturer's instructions. In short, extract mitochondria from ovarian tissue to prepare homogenate. Upon centrifugation, 50 µl Mitochondrial homogenate was mixed with 50 µl catalyst solution and 100 µl (DCFH) solution for 30 min. The 2',7'-dichlorodihydrofluorescein (DCF) levels were quantified using fluorescence spectrometry with excitation/emission (Ex/Em) at 480/530 nm.

Enzyme-linked immunosorbent assay

In short, weigh 0.1 g of ovarian tissue, add cold phosphate buffered saline at a ratio of 1:9, and prepare 10% ovarian tissue homogenate, centrifuged at 14,000 × g at 4 °C for 5 min, and collected supernatants. MDA, GSH, 8-OHdG was determined by a colorimetric assay kit (F2221-A, F30753-A, F2658-A, Shanghai Fanke Industrial Co., Ltd, Shanghai, China) according to the manufacturer's guidelines.

Transmission electron microscope

The ovaries were collected and fixed in Transmission Electron Microscope (TEM) (Servicebio, China) at 4 °C, and dehydrated with gradient concentrations of ethanol. Then, the ovaries were penetrated and embedded with resin and sliced. Then, the ovary section was stained with uranium acetate and lead citrate. Finally, the ovary section was observed under TEM (Hitachi, Japan), and images were captured.

Immunofluorescence staining

In brief, Paraformaldehyde-fixed ovaries were dehydrated, embedded, and sliced. Primary antibodies against PINK1 (proteintech, 23274-1-AP, 1:200) and Parkin (proteintech, 66674-1-Ig, 1:50) were added at 4 °C overnight, while the secondary antibodies were added to the sample at 37 °C for 30 min (Parkin: Servicebio, GB22301, 1:100; Pink1: Servicebio, GB21303, 1:200). DAPI was added at room temperature for 10 min. Images were collected by section using a fluorescence microscope camera system and analyzed by Image J software, randomly select areas of the same size from each slice to measure fluorescence intensity, with three replicates per group.

Prussian blue staining

Prussian blue staining kit (Solarbio, China) was used to detect the deposition of iron-containing heme in ovarian tissue. The paraffin sections were prepared with a thickness of 4 µm, subjected to routine dehydration and transparency, and then stained with Perls staining solution for 30 min. The slices were thoroughly washed with distilled water for 30–50 s, and the background was lightly stained with eosin staining solution for 15–30 s, followed by rinsing with distilled water for 2–3 s. Finally, the slices were dehydrated with anhydrous ethanol, transparent with xylene, sealed with neutral glue, and placed under a light microscope for observation.

Iron assay

In short, ovarian tissue was weighed and homogenized in 1 ml ice-cold phosphate-buffered saline, centrifuged at 14,000 × g at 4 °C for 5 min, and collected supernatants. The total iron content in the ovary were detected by the Total Iron Colorimetric Assay Kit (Elabscience, E-BC-K772-M). Spectrophotometry was finally adopted to detect the absorbance at a wavelength of 593 nm.

Western blot analysis

The expression levels of mitophagy-related proteins PINK1, Parkin and ferroptosis-related proteins GPX4 and ACSL4 were detected by Western blot. After adding radioimmunoprecipitation assay lysis buffer (Beyotime), tissues were ground in a homogenizer and then lysed on ice for 1 h. The Protein concentration was determined using the BCA kit (Solarbio, Beijing, China). The protein loading buffer was added to each sample which was then boiled for 10 min, centrifuged at 4 °C for 10 min and stored. Following protein gel electrophoresis, membrane transfer for 1 h at a constant current of 250 mA in a 4 °C transfer solution, blocking, and incubation of primary antibody and secondary antibody, the photographic developer was added to the membrane and the band was developed in ECL chemiluminescence instrument with

glyceraldehyde-3-phosphate dehydrogenase as internal reference. The protein bands were analyzed by Image J software (version: v1.8.0). The antibodies as follows: anti-GPX4(Abcam; ab125066; 1:5000), anti-ACSL4(Abcam; ab155282; 1:10000), anti-Parkin(Abcam; ab77924; 1:1000), anti-PINK1(Abcam; ab300623; 1:1000), anti-GAPDH(Proteintech; 60004-1-Ig; 1:5000).

Statistical analysis

Continuous variables are expressed as mean±SD throughout this study. Differences between the two groups were analyzed using one-way ANOVA followed by Duncan's post hoc test. All statistical analyses were performed with the SPSS 25.0 and Image-J statistical software. A P -value<0.05 was considered statistically significant.

Results

EZTG improves ovarian function in aged mice

The ovaries of the young group are well-developed, and follicles at all grades are visible. Compared with the young group, the old group showed a decrease in the number of corpus luteum in the ovaries, a loose arrangement of granulosa cells, and a decrease in the ovarian index (ovarian/body weight) (Fig. 1A, C). According to the follicle count results, the primordial, primary, secondary, and mature follicles in the EZTG-H group were significantly higher than those in the old group ($P<0.05$) (Fig. 1B). Compared with the young group, the serum AMH levels in the old group were significantly reduced ($P<0.01$), while FSH levels were increased. After intervention with medium and high doses of EZTG, compared with the old group, the levels of AMH significantly increased ($P<0.01$), while the levels of FSH decreased ($P<0.05$) (Fig. 1D-E).

EZTG improves mitochondrial function in the ovaries of aged mice

In order to explore the effect of EZTG on mitochondrial dysfunction, we used JC-1 and flow cytometry to study the level of MMP in the ovary. Compared with the young group, the MMP in the old group was significantly lower. However, in EZTG-L and EZTG-H groups, the MMP increased significantly compared with the old group ($P<0.05$) (Fig. 2A,B). At the same time, we measured the levels of markers of mitochondrial oxidative damage 8-OHdG and mitochondrial ROS. Compared with the young group, the 8-OHdG and mitochondrial ROS in the old group were significantly increased. In the EZTG-L and EZTG-H groups, 8-OHdG and mitochondrial ROS were significantly decreased compared to the old group ($P<0.05$)(Fig. 2C, D). The mitochondrial ultrastructure of ovarian tissue was observed using TEM. In the young group, mitochondrial morphology is intact, with more

MDVs. The number of mitochondria in the old group decreased, with fragmented morphology and less MDVs. However, the administration of EZTG maintained mitochondrial homeostasis (Fig. 2E).

EZTG improves the level of mitophagy in the ovaries of aged mice

To detect mitophagy levels, we used immunofluorescence staining to measure the expression of Parkin and PINK1 in the ovaries and measured protein expression using Western blot. Compared with the young group, the fluorescence expression levels of PINK and Parkin in the old group were significantly reduced. The fluorescence expression levels in the EZTG-L and EZTG-H groups were significantly higher than in the old group (Fig. 3A, B, D, E). Similarly, the results of Western blot also showed a similar trend (Fig. 3C, F, G).

EZTG can alleviate ferroptosis in the ovaries of aged mice

Prussian blue staining was used to evaluate iron deposition in ovarian tissue. The results showed a significant increase in iron deposition in the old group compared to the young group. The iron deposition in the EZTG-L and EZTG-H groups was significantly reduced compared to the old group (Fig. 4A). Similarly, there is a similar trend in the levels of Iron ions in the ovaries (Fig. 4C). TEM observed the mitochondrial morphological changes showed that the mitochondria in the old group were smaller than those in the young group, the mitochondrial membrane density was condensed, mitochondria crista was reduced, and the outer mitochondrial membrane was ruptured. The mitochondrial morphology of the ovaries in both EZTG-L and EZTG-H groups improved (Fig. 4B). Lipid peroxidation is one of the main mechanisms of Ferroptosis. We evaluated the levels of GSH and MDA in ovarian tissue. Compared to the young group, the GSH level in the old group decreased while the MDA level increased. The GSH levels in the EZTG-L and EZTG-H groups increased, while MDA levels decreased compared to the old group (Fig. 4D, E). The protein blotting results showed a decrease in GPX4 levels in the old group, while an increase in ACSL4 levels was observed compared to the young group. The levels of GPX4 increased in the EZTG-L and EZTG-H groups, while the levels of ACSL4 decreased compared to the old group (Fig. 4F, G, H).

Discussion

Ovarian aging in old women is the main cause of infertility and an intractable clinical issue. Whether using the customized IVF-ET regimen [17] or the combination of other drugs, such as Dehydroepiandrosterone, Luteinizing hormone [18], improving the clinical pregnancy outcome of AMA is challenging. This study established a mouse model of natural aging to study the effect of EZTG

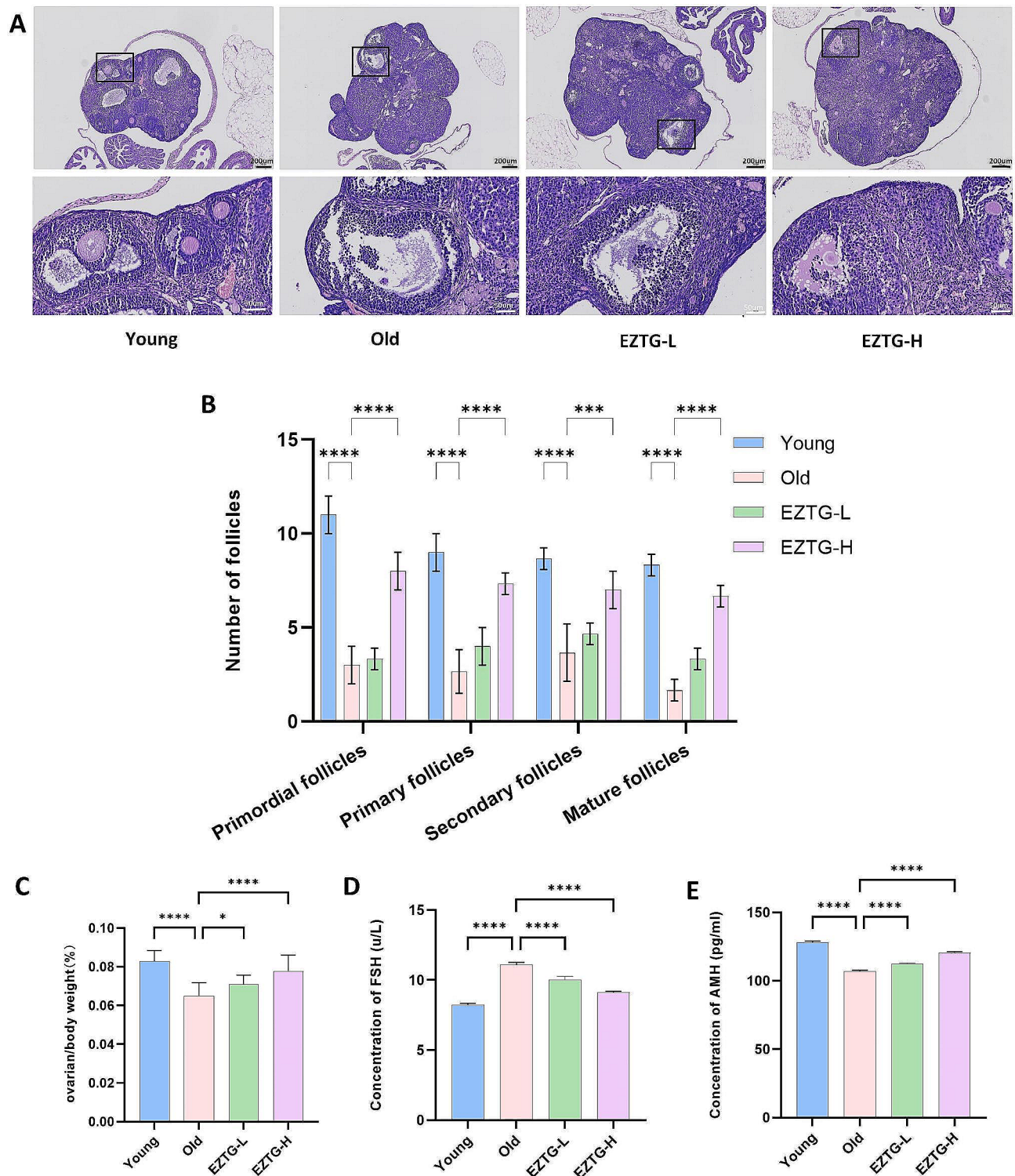


Fig. 1 EZTG improves ovarian function in aged mice. **(A)** Ovarian tissue sections with H&E staining in different groups. **(B)** Follicle count at all grades. **(C)** ovarian index (ovarian/body weight). **(D,E)** The level of serum follicle stimulating hormone anti-Müllerian hormone. * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$, **** $P < 0.0001$. All data are expressed as the mean \pm SD and at least three independent experiments were performed

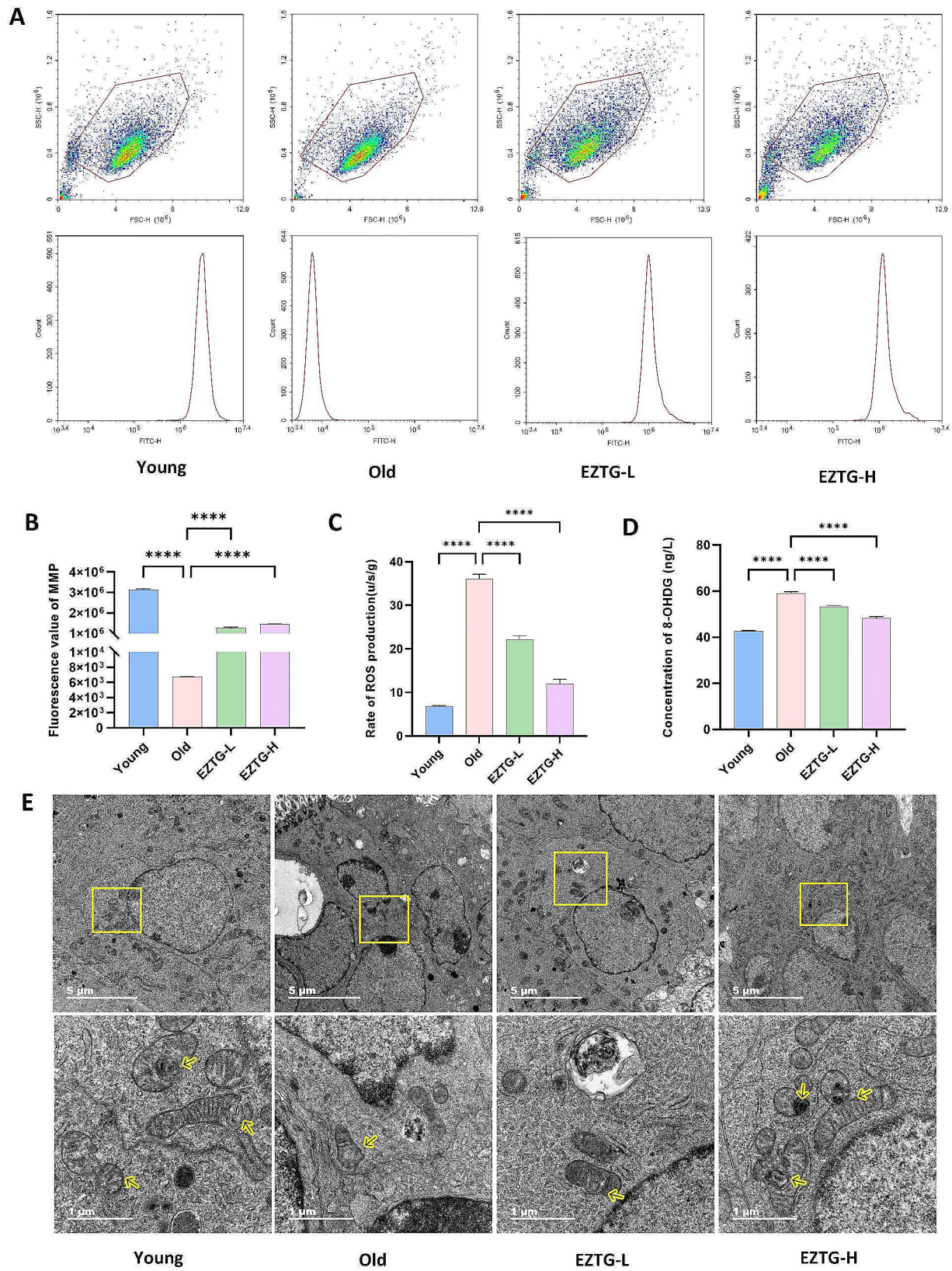


Fig. 2 E2TG improves mitochondrial function in the ovaries of aged mice. **(A,B)** Flow cytometric analysis of MMPs degradation. **(C)** Rate of Mitochondrial ROS production **(D)** concentration of 8-OHdG. **(E)** Observation of mitochondrial ultrastructure and MDVs in ovaries under transmission electron microscopy, the yellow arrow identifies MDVs, scale bar + = 5 μm. **P* < 0.05, ***P* < 0.01, ****P* < 0.001, *****P* < 0.0001, All data are expressed as the mean ± SD and at least three independent experiments were performed

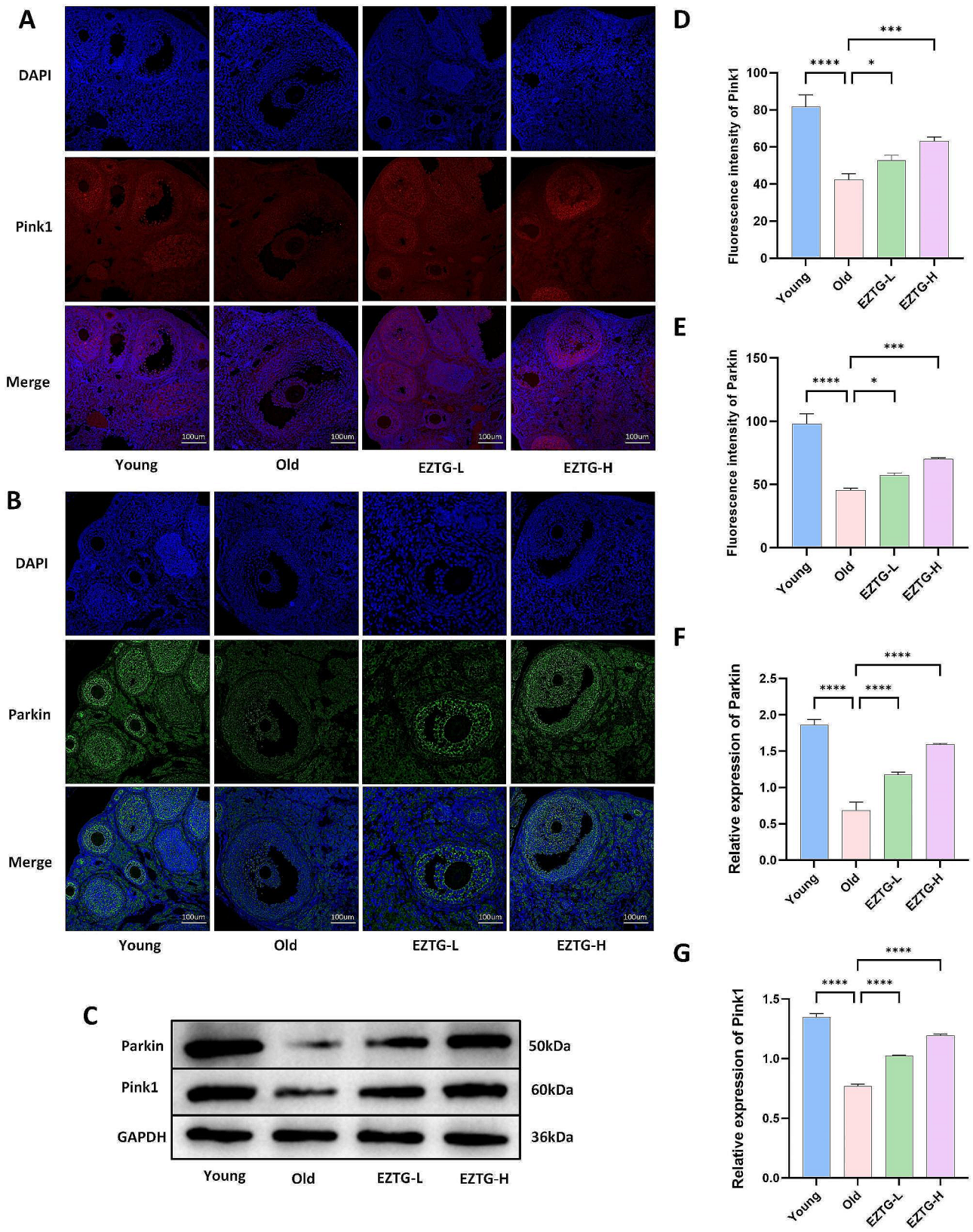


Fig. 3 EZTG improves the level of mitophagy in the ovaries of aged mice. **(A,B)** Immunofluorescence images of the PINK1 and Parkin. **(C)** Representative protein bands of PINK1 and Parkin. **(D,E)** Quantification of Immunofluorescence staining for PINK1 and Parkin. **(F,G)** The corresponding quantitative statistical results of PINK1 and Parkin protein. $*P < 0.05$, $**P < 0.01$, $***P < 0.001$, $****P < 0.0001$, All data are expressed as the mean \pm SD and at least three independent experiments were performed

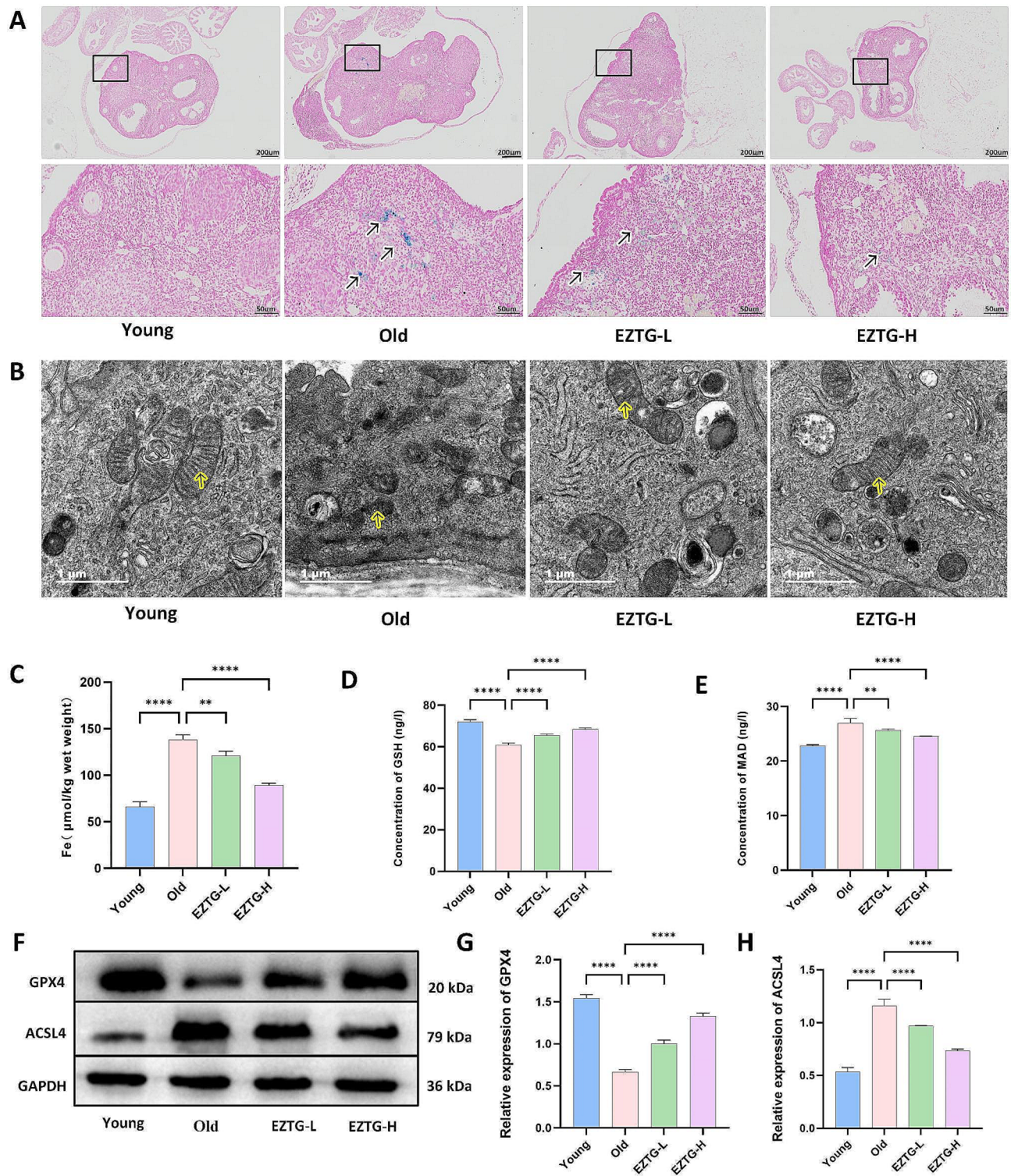


Fig. 4 EZTG can alleviate ferroptosis in the ovaries of aged mice. **(A)** Ovarian tissue sections with Prussian blue staining, scale bar += 50 µm. **(B)** Observation of Mitochondrial morphology in ovaries under transmission electron microscopy, the yellow arrow identifies mitochondrial cristae, scale bar += 1 µm. **(C.D.E)** Quantitative analysis of iron ions, GSH and MDA in ovaries. **(F)** Representative protein bands of GPX4 and ACSL4. **(G.H)** The corresponding quantitative statistical results of GPX4 and ACSL4 protein. * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$, **** $P < 0.0001$, All data are expressed as the mean ± SD and at least three independent experiments were performed

on alleviating age-related ovarian aging. The experimental results indicate that EZTG can improve ovarian endocrine function and increase the number of follicles. In addition, EZTG can maintain PINK1/Parkin-mediated mitochondrial homeostasis, reduce the lipid peroxidation caused by the accumulation of ROS, and inhibit the occurrence of ferroptosis and delaying ovarian aging.

Studies based on clinical samples reveal that Erzhi-Tiangui Granule alleviates ovarian aging and its effect on follicular fluid and granulosa cells in AMA. EZTG can improve embryo quality in elderly infertile women with kidney Qi and Yin deficiency syndromes and reduce the apoptotic effect in the follicular fluid [19]. Meanwhile, EZTG increased oocyte fertilisation rate and affected granulosa cells' mRNA expression profile in patients with premature ovarian failure [20]. In this study, we observed that aged mice showed increased ovarian reserve, improved serum hormone levels after EZTG treatment. Many famous traditional Chinese medicines in EZTG include *Lycium chinense* Mill, *Angelica sinensis*, *Paeonia lactiflora* Pall, etc. *Lycium chinense* Mill can repair ovarian injuries by scavenging oxidative products 8-hydroxy-2'-deoxyguanosine and increasing anti-mullerian hormone expression in mice with ovarian injuries induced by repeated superovulation [21]. *Angelica sinensis*, also known as "female ginseng", is renowned for treating various gynaecological conditions [22]. *Angelica sinensis* extract could enhance the invasion and migration ability of follicle micro-vascular endothelial-like cells of chicken *in vitro* and promote the proliferation of granulosa cells [23]. *Paeonia lactiflora* Pall has long been used to treat female infertility and recurrent miscarriages. The water-extract *Paeonia lactiflora* Pall increases ovarian expression of angiogenic factors and follicular development regulatory factors in aged female mice [24].

Mitochondrial dysfunction induces ovarian aging, not only perturbs oocyte-cumulus crosstalk in the ovary [5] but also affects the processes during oocyte meiosis and the developmental potential of embryos [25]. Mitochondrial dysfunction is reflected in many aspects, including the decrease of MMP, the increase of mtDNA mutations, and the accumulation of ROS, etc [26, 27]. In our study, we observed the decrease of MMP, the increase of ROS and 8-OHdG in the ovary of aging mice, indicating mitochondrial dysfunction. After further administration of EZTG, mitochondrial dysfunction was alleviated at these levels. The decrease of MMP, considered a sign of mitochondrial dysfunction, is typically associated with increased production of ROS [28, 29]. Mitochondria are the main source of free radicals and ROS in cells [30]. The mitochondrial disorder produces excessive ROS, and a considerable accumulation of ROS leads to oxidative stress, which in turn affects mitochondrial homeostasis and forms a vicious cycle [6, 7]. In elderly women

receiving assisted reproductive therapy, the concentration of ROS in their oocytes, cumulus oophorus and follicular fluid increase [31], while antioxidant decreases [32]. The mtDNA is vulnerable to ROS attack due to lack of histone protection and overlaps with ROS production sites in the Inner mitochondrial membrane [33], leading to mitochondrial dysfunction and premature aging [34]. Among many types of mtDNA oxidative lesions, 8-OHdG is an indicator of steady-state level of oxidative damage occurring within mtDNA [35].

Mitophagy is a critical mitochondrial quality control mechanism that maintains mitochondrial homeostasis by selectively eliminating damaged or excessive mitochondria [36]. Mitophagy activity decreases with age, and its upregulation may improve symptoms of age-related phenotypes [37]. Our study found that the levels of PINK1 and Parkin in the ovaries of aged mice decreased, indicating a decrease in mitophagy levels. At the same time, we observed changes in MDVs in the ovaries of aged mice under electron microscopy, EZTG reversed this impact to some extent. PINK1/Parkin mainly regulates ubiquitin-dependent mitophagy, which is one of the main mechanisms of autophagy and is related to its initiation [38]. Knockout of PINK1 in *Drosophila* leads to infertility and mitochondrial dysfunction, which can be alleviated by overexpression of Parkin [39]. In addition to mitophagy, Parkin and PINK1 have been shown to control mitochondrial quality through a mitophagy-independent mechanism of mitochondria turnover via the formation of MDVs [40].

Mitochondria regulate various forms of cell death and play a crucial role in the regulation of ferroptosis [8, 9]. The morphological changes of mitochondria, including reduced mitochondrial volume, increased bilayer membrane density and reduction or disappearance of mitochondrial cristae, are also one of the signs of ferroptosis. Ferroptosis, a novel form of regulated cell death driven by iron-dependent lipid peroxidation, is characterized by the increase of free iron and accumulated lipid peroxides, which produces intracellular ROS [41]. Iron ion, a major indicator of ferroptosis, oxidizes lipids in a Fenton-like manner, thus promoting ferroptosis [42]. Depleting intracellular GSH leads to the decrease of GPX4 activity, leading to the disorder of lipid oxide metabolism, thus promoting ferroptosis. Among them, MDA is one of the products of Lipid peroxidation [43]. GPX4, at the crossroads of lipid homeostasis and ferroptosis, the functional inactivation of GPX4 results in an increased lipid-ROS accumulation and subsequent lipid peroxidation [44]. Meanwhile, ACSL4 promotes sensitivity to ferroptosis by shaping cellular lipid components [45]. In our study, we observed the aggregation of iron ions, the increase of Lipid peroxidation products, the decrease of GSH and GPX4, and the increase of ACSL4 in the ovaries

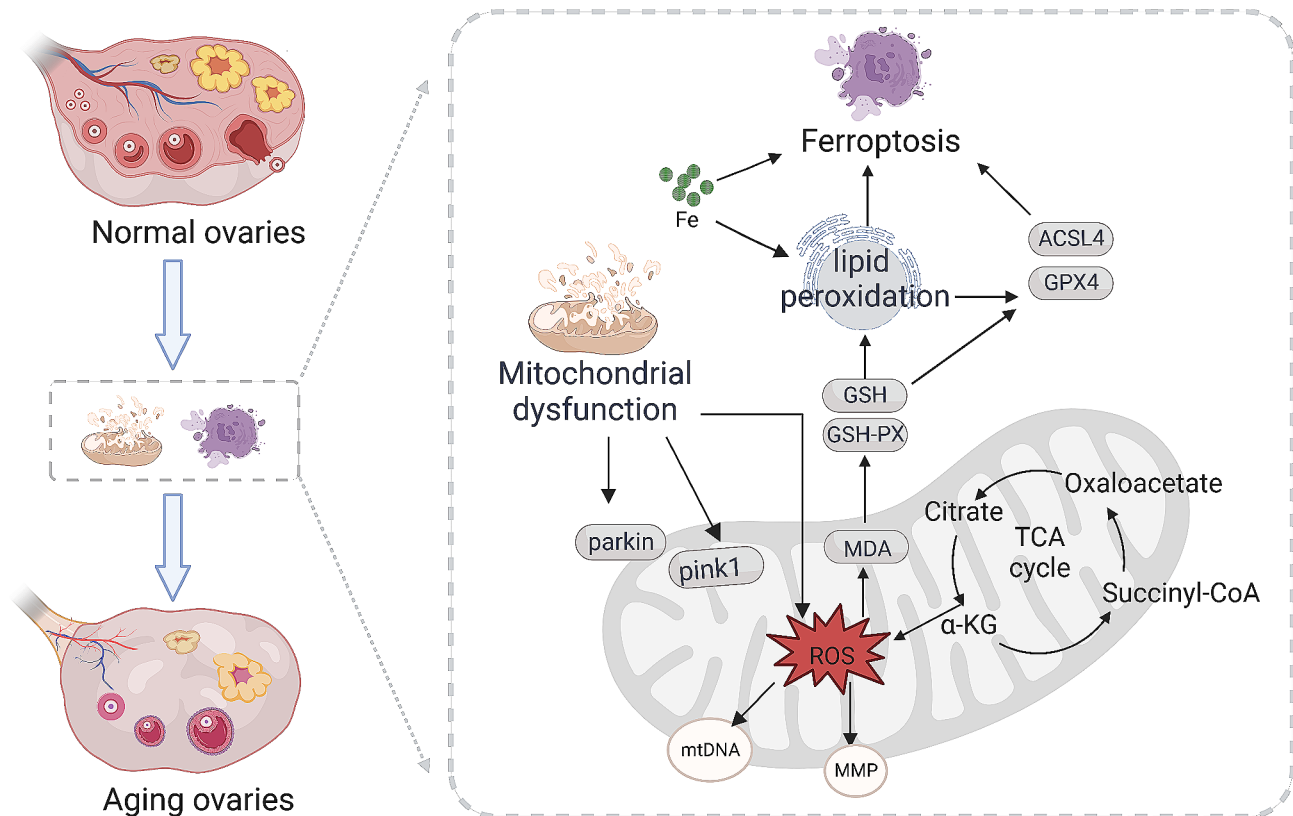


Fig. 5 Schematic illustration showing that EZTG alleviates age-related ovarian aging by regulating mitochondrial homeostasis to inhibit ferroptosis

of aged mice. Similarly, we observed reduced mitochondrial volume, increased bilayer membership density and reduction or disappearance of mitochondrial cristae in the ovaries of aged mice. This indicates that ferroptosis occurs in age-related ovarian aging. EZTG alleviated the occurrence of ferroptosis to varying degrees.

However, our study had some limitations, including a lack of *in vitro* validation experiments to confirm the relationship between mitochondrial homeostasis and inhibiting ferroptosis in ovarian aging. In addition, due to the disordered estrous cycle of elderly mice, on the day of mouse euthanasia, not all mice were in the same estrous cycle stage, which may lead to deviations in corpus luteum quantity and blood hormone levels. At present, almost all relevant animal experimental studies have yet to mention how to control this variable, which is also an idea that we need to advance continuously.

Conclusions

In addition, EZTG can maintain PINK1/Parkin-mediated mitochondrial homeostasis, reduce the lipid peroxidation caused by the accumulation of ROS, and inhibit the occurrence of ferroptosis and delaying ovarian aging (Fig. 5). These findings suggest that EZTG may be a promising drug for treating age-related ovarian aging in females.

Author contributions

JZC performed most experiments, drafted and wrote the manuscript. LYQ, and WPX constructed the animal model. YK and SMY helped revise the manuscript and produce figures. CW and LQH were responsible for processing and analyzing experimental data. GY designed the overall experiment and provided financial support. All authors have contributed to the article and approved the submitted version.

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Data availability

The raw data supporting the conclusions of this article will be made available by the authors, without undue reservation.

Declarations

Ethics statement

Animal Ethics Committee of Affiliated Hospital of Shandong University of Traditional Chinese Medicine approved all the experimental procedures used in this study (Approval number: 20220316).

Competing interests

The authors declare no competing interests.

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