# **CASE REPORT**

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# Controlled ovarian stimulation in cancer patients under 18 years old; a case series



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# Abstract

**Background** Fertility preservation for adolescent pubescent girls is a concern of the healthcare system and parents. Oocyte cryopreservation is regarded as a standard medical intervention for patients with a minimum age of 18 years. Evidence suggests that mature oocyte cryopreservation is possible for adolescent pubescent girls, although, ovarian stimulation for these patients remains a challenge.

**Cases presentation** This case series is the first report regarding ovarian stimulation with oocyte cryopreservation in younger than 18 years cancerous girls, who refer to ROYAN institute, Tehran, Iran, prior to the start of the treatment of cancer (November 2015 to February 2021). The oocyte cryopreservation was carried out in the 7 patients (five patients with Hodgkin lymphoma, one patient with Ewing sarcoma, and one patient with osteogenic tumor), the embryo cryopreservation in one patient with dysgerminoma, and the oocyte and embryo cryopreservation in one patient with Hodgkin lymphoma. For one of the patients with Hodgkin lymphoma, half of the tissues of one ovary were cryopreserved prior to ovarian stimulation.

**Conclusions** Oocyte cryopreservation is a feasible option of fertility preservation in the adolescent's patients with cancer. However, only if reported acceptable fertilization rates, as well as the successful cases of live birth from oocyte cryopreservation at the ages under 18, this option of preserving fertility can be applied to this age range.

**Keywords** Fertility preservation, Oocyte cryopreservation, Controlled ovarian stimulation, Cancer patients, Adolescent pubescent girls, Case series

# Background

Although the survival of children and adolescents with cancer has significantly increased in recent decades, cancer survivors suffer from long-term complications of this disease or its therapies, such as gonadotoxicity of therapies. The importance of enhancing this patient's quality of life and preserving their ability to have a

<sup>1</sup> Department of Endocrinology and Female Infertility, Reproductive Biomedicine Research Center, Royan Institute for Reproductive Biomedicine, ACECR, Number 12, East Hafez Avenue, Banihashem Street, Resalat Highway, Tehran, Iran biological child in the future life highlights the necessity of fertility preservation (FP) strategies. Preserving the fertility of adolescent pubescent girls is a hot topic of discussion [1, 2].

The main options for FP in cancer patients include mature oocyte, or embryo, and ovarian tissue cryopreservation (OTC). Mature oocyte or embryo cryopreservation (EC) is a standard FP method in post-pubescent women if they have had adequate time for ovarian stimulation (OS). OTC is a FP method for pre-pubescent girls and post-pubescent patients who cannot delay the onset of cancer treatment [3, 4]. Previously, it was considered experimental; however, the American Society for Reproductive Medicine (ASRM) proposed OTC as an established method [5]. Although pediatric research must



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continue [6]. This technique is more invasive than other FP methods as it needs laparoscopy and, if necessary, laparotomy to remove and re-graft the ovarian tissue; there is also the risk of tissue contamination with cancer cells and the transmission of the disease following tissue graft [3, 4].

Recent advances in the cryobiology significantly promoted oocyte cryopreservation (OC), while lack of sperm dependence has made this method a desirable option. OC is regarded as a standard medical intervention for patients with a minimum age of 18 years, although several studies introduced it enough safe to use for younger patients [7–17].

Although evidence suggests that mature OC is possible for adolescent pubescent girls, it is controversial due to the dearth of research on OS in these patients. The controversial topics related to OS and mature OC in the cancer affected adolescents that included the unknown success rate of cryopreserved oocyte usage in this age group [7]; no experience of sexual intercourse, which may cause psychological problems for them [17]; parent decision-making on behalf of the patient; inadequate knowledge about adolescents' response to OS; adolescents finding it difficult to accept this process due to the necessity of daily injections, ultrasound, and a series of lab tests [18]; the possible immaturity of the hypothalamus-pituitary-ovarian (HPO) axis that may lead to an inappropriate ovarian response; the unreliability of factors that determine the prescribed gonadotropin dose for OS in this age group, including anti-mullerian hormone (AMH) and antral follicle count (AFC) [10, 19]; and inherent differences in the adolescent ovary physiology, especially in the first postpubescent years [17].

The present study reports cases of OS in cancer patients younger than 18 years in ROYAN institute, Tehran, Iran (2015-2021). As well as, reporting the outcomes of OS cycles for OC, the associated challenges in this age group are also discussed.

# **Cases presentation**

This case series study was approved by the Research Ethics Committee of ROYAN institute, Tehran, Iran (Ethics Approval Code: IR.ACECR.ROYAN.REC.1400.080). Between November 2015 and February 2021, the patients of our Oncofertility and FP Clinic, Royan institute, Tehran, Iran, who have the inclusion criteria, were invited in this study. Our main criteria were included (1): younger than 18 years (2): refer before cancer therapies. The process of admission oncofertility cases in this center is presented in Fig. 1.

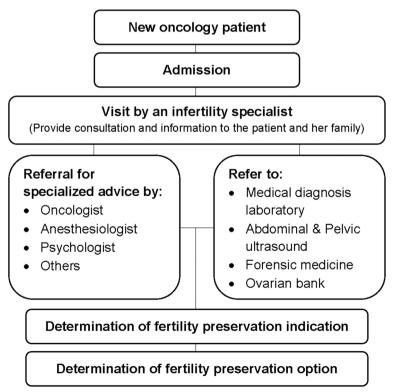


Fig. 1 The process of admission oncofertility cases in ROYAN Institute

Ten patients between the ages of 14 and 17, and 1 to 6 years from their Menarche age (the first menstrual cycle) entered the cycle of the OS. These patients suffered from Hodgkin Lymphoma (HL) (n=5), ovarian tumor (germ cell tumor and dysgerminoma) (n=2), Ewing sarcoma (n=1), osteogenic tumor (n=1) and brain tumor (medulloblastoma) (n=1). The OS was performed for all 10 patients using the standard gonadotropin releasing hormone (GnRH) antagonist protocol. The OS beginning was different among patients: in the early proliferative phase (n=8), during the late proliferative phase (n=1), and in the luteal phase (n=1).

Trans-abdominal ultrasound was used successfully for follicular monitoring. The gonadotropin prescribed for all patients was recombinant follicle stimulating hormone (r-FSH) (Gonal-f) (Follitropin alfa, Merck Serono, Germany): for eight patients began with a dose of 150 IU/ day, for the patient in the luteal phase with a dose of 225 IU/day, and for the patient with medulloblastoma with a dose of 112.5 IU/day. The prescribed drug for triggering final oocyte maturation was recombinant human chorionic gonadotropin (r-HCG) (500  $\mu$ g Ovitrelle) (Choriogonadotropin alfa, Merck Serono, Switzerland) in the patients who underwent OS during 2015-2018 and lowdose GnRH-agonist (0.2 mg triptorelin) (Decapeptyl, Ferring Pharmaceuticals, Germany) for the other patients.

The OC was carried out in 7 patients, the EC in one patient and the oocyte and embryo cryopreservation in one patient. No oocytes were retrieved after OS in the patient with medulloblastoma. For one of the patients with HL, half of the tissues of one ovary were cryopreserved prior to OS. The two patients with ovarian tumor had a history of unilateral oophorectomy.

Baseline characteristics of the patients are presented in Table 1. Information of patients' OS cycle and its results is presented in Table 2. As well as, the summary of similar studies is presented in Table 3.

# **Discussion and conclusions**

According to American Society of Clinical Oncology (ASCO) guideline all newly diagnosed cancer patients of reproductive age should be informed about potential loss of fertility and should refer to infertility specialists [20]. Recommendation of gamete and/or embryo cryopreservation before gonadotoxic treatments is currently a standard of care [21]. FP in the adolescent patients is very rare and typically performed before gonadotoxic therapies for oncologic or non-oncologic conditions, such as a stem cell transplantation, or impending premature ovarian failure (POF) [7].

There are many dilemmas about FP among adolescent populations, including parental decision-making on behalf of the patient, adolescent decision capacity [22], and loss of virginity due to the trans-vaginal approach during oocyte retrieval. The latest one, loss of virginity is a too difficult condition for the parents, particularly in some cultural perspectives [10].

According to the instruction followed in our center, talking to adolescents about FP, including its necessity and steps, is done by a fertility specialist in the presence of their parents. The specialist explains these phases' step by step in a language understandable to them. Their questions are answered. If the adolescent and her parents' consent, the parents are asked to sign the consent forms. Given the importance of virginity in Iranian Culture, our center adheres to the following instruction to prevent injuring the hymen during follicle puncture. After applying anesthesia the patient put in the lithotomy position, the infertility specialist separates the labia minora, making the hymen more visualized. Lubricates her little finger, and gently touches the edges of the hymen. And tries to dilate hymen when the little finger entered the vagina. Infertility specialist attempts to enter the ring finger and continue to dilate hymen and when the ring finger freely moves the physician tries middle finger and after that index and then thump and finally both index and middle fingers tries simultaneously. The procedure takes 10-15 minutes to perform. Now, a suitable speculum can be used without injuring the hymen. To eliminate the lubricant effects, the vaginal environment is washed with vigorous amount of normal saline and dried with a small sterile gauze before the insertion of the ultrasound probe. The remaining steps of follicle puncture will be similar to adults' instructions. In this study out of patients were virgin. And all of the intact hymen were annular shape. And none of them were injured during follicular puncture.

Knowledge gaps exist as to whether adolescence patients respond similarly to the OS or have similar outcomes, such as mature oocyte yield. Currently, published data in adolescents' are limited to case series [10, 14, 23, 24]. Previous studies in the FP for oncology patients have shown lower oocyte yield in adult patients [25, 26]. Studies indicated the cancer pathology might itself induce ovarian reserve depletion [25]. As well as, cancer can induce an increased catabolic condition, increase in stress hormones with associated hypothalamic dysfunction; can predispose patients to lower levels of gonadotropins [18]. There are few studies that detail response to the OS and some of the studies had raised concern that adolescent patients might have a lower response to gonadotropins [7].

Although, sensitive markers of ovarian reserve, including AFC and AMH in estimating the dose of gonadotropins likely to yield maximum oocyte have been well documented, there is limited data of dose specific

Case	<del>,</del>	2	£	4	5	6	7	80	6	10
Year	2015	2017		2018		2019			2020	2021
Diagnosis	HΓc	Ovarian tumor (Germ cell) HL	Η	НГ	Ewing Sarcoma	Ewing Sarcoma Ovarian tumor (Dysgerminoma)	H	Brain tumor (Medulloblastoma)	Osteogenic tumor	Η
Age (year)	16	16	17	17	16	15	17	16	14	17
Menarche age	12	13	N/A	11	10	13	14	12	13	;;
<b>BMI</b> (kg/m <sup>2</sup> )	19.1	19.9	18.1	20.3	19.2	19.5	19.6	19.1	24.9	19.1
Ovarian surgery <sup>a</sup>	<b>Ovarian surgery</b> <sup>a</sup> Cryopreservation of <sup>1</sup> / <sub>2</sub> left ovarian tissue <sup>d</sup>	Right Oophorectomy	No	No	No	Right Oophorectomy	No	N	No	No
Radiotherapy <sup>b</sup>	No	No	No	No	No	No	No	No	No	No
<b>Chemotherapy</b> <sup>b</sup>	No	No	No	No	No	No	No	No		No
Menstruation	Regular	Irregular	Irregular	Irregular	Regular	Irregular	Regular	Irregular	Irregular	Regular
AFC	4	9	16	14	16	9	14	20		20
AMH (ng/ml)	N/A <sup>e</sup>	0.4	6.0	0.9	5.1	1.9	N/A	8.9	N/A	6.0
FSH (IU/L)	3.7	4.2	9.1	9.1	3.4	6.7	4.5	3.9	0.0	2.2
<b>LH</b> (IU/L)	9.4	8.8	7.8	5.3	2.6	12.4	3.2		12.5	5.0
Prog (ng/ml)	0.2	2.5	0.2	1.1	N/A	0.5	N/A	0.2	0.2	1.2
<b>E2</b> (pg/ml)	88.3	141.9	40.3	188.0	N/A	33.2	N/A	17.5	144	N/A

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 $^{\mathrm{b}}$  Radiotherapy and/ or chemotherapy history before ovarian stimulation

<sup>c</sup> Hodgkin lymphoma

 $^{\rm d}$  Cryopreservation of ovarian tissue with the aim of fertility preservation  $^{\rm e}$  Not Available

Case	1	2	3	4	5	6	7	8	9	10
COS Protocol	GnRH anta	agonist								
Stimulation cycle start	Early. P <sup>b</sup>	Luteal Phase <sup>c</sup>	Early. P <sup>d</sup>	Late. P <sup>e</sup>	Early. P					
Gonadotropins	recombin	ant-FSH								
Starting gonadotropin dose (IU)	150	225	150	150	150	150	150	112.5	150	150
Total gonadotropin dose (IU)	1050	2025	1350	1500	1200	1650	1425	1687.5	1350	1350
GnRH-antagonist (ampule)	4	4	5	6	4	8	3	9	6	5
Duration of stimulation (day)	8	10	10	11	9	12	9	14	10	11
Follicle> 12 mm <sup>a</sup>	10	6	12	11	13	17	11	13	8	22
Triggering medication	2recombii	nant-HCG				Low do	ose GnRH-	agonist		
Total oocytes retrieved	13	10	9	10	21	6	8	0	32	33
Total mature oocytes (MII)	13	10	9	8	17	5	5	0	26	23
Total oocytes cryopreservation	13	5	9	8	20 <sup>f</sup>	0	7 <sup>f</sup>	0	26	23
Total embryos cryopreservation	-	5	-	-	-	5	-	-	-	-
OHSS & others side effects	No									

Table 2 Information of patients' ovarian stimulation cycle and its results

<sup>a</sup> In the day of triggering final oocyte maturation

<sup>b</sup> Proliferative

<sup>c</sup> Progesterone >1.5 ng/ml

<sup>d</sup> Larges follicle <13 mm & progesterone <1.5 ng/ml

<sup>e</sup> Larges follicle >13 mm & progesterone <1.5 ng/ml

<sup>f</sup> MII+GV (germinal vesicle)

evaluation in pubertal girls [27]. As well as, there are arguments regarding the clinical value of AMH assessment in adolescent girls [28]. In our study, the correlation between AFC, AMH, gonadotropin dose, and the number of oocytes retrieved was not statistically significant, which could be partially attributed to the small sample size. Accordingly, judging the value of AFC and AMH for determining the gonadotropin dose and number of oocytes retrieved in the adolescent population requires a study with a larger sample size. Antral follicles are present in the ovaries of girls of all ages. At the onset of puberty, a rise in the FSH levels happens and antral follicles start growing. Primary menstruation cycles are frequently anovulatory, with regular ovulation occur later. While maturity of the hypothalamus-pituitary-axis is crucial for puberty, changes in ovarian maturity may also contribute. This may be reflected with little but constant transient reduction in the AMH level during puberty. There are significant numbers of follicles with abnormal morphology during puberty, these seem to be lost during adolescence. Follicle growth may also be different in their ovaries. Also, isolated secondary follicles from adolescents grew slower than those of adults that indicates an inherent maturity change. Although, its basis has remained unknown. Moreover, differences in local regulatory factors, that reflects the high density of small follicles in these young patients, may also contribute to the pattern of ovarian development. A discrepancy has been found among AFC, AMH and number of oocytes cryopreserved in some cases. These differences are likely multifactorial and could be a reflection of the ovarian development stage at extremes of youth or attributed to the effect of the chronic illness [10].

Although studies have suggested that trans-abdominal ultrasound may decrease the accuracy of AFC, overall this did not pose a significant issue in the adolescent treatment and had previously been successfully used for monitoring in the similar populations [10, 15–17]. In our study, the condition of adolescents' ovaries, including AFC, was examined by a skilled radiologist through the abdomen before the onset of the OS.

There are few studies in the adolescent patients that detail complications, delay in the initiating cancer treatment, and pregnancies after the completion of treatment. Most patients with cancer diagnosis can be afforded the 8-12 days required for the OS before they begin their treatment, without imperiling their oncologic care [17]. In our study, the OS delayed the adolescent cancer treatment by 10-14 days. It is worth noting that the cancer treatment delay was done after receiving confirmation from the oncologist. No complication was reported as a result of delayed cancer treatment.

The GnRH-antagonist protocol has decreased the interval between patient presenting and OS. The shorter duration of treatment, followed by a minimally invasive trans-vaginal oocyte retrieval, with a short recovery period, compared with laparoscopy, makes the procedure more acceptable to patients [10]. The use of

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Author; Year	Kutteł	Kutteh et al, 2018 [9]	018 [9]	Peddie et al, 2018 [ <b>16</b> ]		Lavery et al, 2016 [10]	016 [10]	l					Oktay	Oktay et al, 2014 [14]				Reichman et al, 2012 [ <mark>17</mark> ]	Kim et al, 2011 [8]	Oktay et al, 2010 [15]	Noyes et al, 2009 [13]	Nagashima et al, 2005 [12]	ima 05
Patients	m	2	Ļ	-	00	7	9	5	4	m	2	-	5	4	m	2	-	-	1	1	1	2	-
Diagnosis	Medu	Medulloblastoma	oma	myelo- dys- plastic clone <sup>f</sup>		Sickle cell anemia	mia						ALL	Germ cell tumor	Tumer syn- drome			Myelod- ysplastic syndrome	SPH caused by TGV <sup>d</sup>	Turner syndrome	Ovarian lutein- ized the coma	AML	17
Age (Menarche age)	14 (13)	16 (12)	14 (13)	14 (12)	18	18	17	16	16	16	15	14	14 (13)	15 (13)	14 (11)	13 (12)	13 (13)	13	17	14 (11)	19	18	No
Ovarian surgery	°Z	No	No	No	°Z	°N N	No	N	°N	No	°Z	°Z	°Z	Left oophorec- tomy	N	No	0 N	oz	oN	OZ	Left oophorec- tomy	No	N
Radiotherapy	No	°N N	°N N	No	No	No	°N N	No	°N N	No	No	°N N	°N N	No	No	No	No	No	No	No	No	No	Yes
Chemotherapy	No	No	No	No	No	No	°N N	No	No	No	No	°N N	Yes	No	No	No	No	No	No	No	No	No	N/A <sup>a</sup>
AMH (ng/mL)	12.59	6.8 <sup>9</sup>	13.39	N/A	$16^{e}$	N/A	24 <sup>e</sup>	23.8 <sup>e</sup>	N/A	N/A	N/A	N/A	0.8	1.6	0.9	0.76	1.59	0.95	N/A	0.9	N/A	N/A	N/A
FSH (mIU/mL)	N/A	N/A	N/A	7.1	2.9	7.6	4.2	7.6	4.3	1.2	4.8	2	7.8	5.6	5.3	5.6	5.7	5.0	N/A	5.3	N/A	N/A	N/A
<b>LH</b> (mIU/mL)	N/A	N/A	N/A	10.4	3.2	3.9	3.6	3.8	3.3	0.8	6.2	6.2	8.1	9.2	9.5	5.3	3.9	2.9	N/A	9.5	N/A	N/A	N/A
AFC	31	15	25	17	12	20	20	16	16	18	9	13	5	11	12	9	9	6	N/A	12	N/A	N/A	N/A
COS Protocol	GnRH	GnRH-antagonist	nist	GnRH-antagonist	ntagoni:	st					GnRH- agonist	- st	GnRH	GnRH-antagonist				GnRH- antagonist	GnRH-agonist	GnRH-antagonist	GnRH-agonist	N/A	N/A
Duration of stimulation	10	12	11	N/A	12	10	11	10	10	11	10	14	12	11	10	10	11	N/A	N/A	N/A	N/A	N/A	N/A
Gonadotropins	DMH			DMH	r-FSH								r-FSH i	r-FSH (+HMG or r-LH)	()			DMH	r-FSH (+HMG)	r-FSH (+HMG)	N/A	N/A	N/A
Starting gonadotropin dose (IU)	150	225	150	225	225	187.5	150	150	112.5	112.5	150	150	N/A	N/A	N/A	N/A	N/A	225	N/A	225 (+150) <sup>c</sup> 225 (+150) <sup>b</sup>	N/A	N/A	N/A
Total gonado- tropin dose (IU)	1800	1800	1950	N/A	3075	3075 1875	3350	1500	1462.5	1312.5	1875	2625	1775	2137.5	2075	2250	2625	N/A	N/A	N/A N/A	N/A	N/A	N/A
<b>Triggering</b> medication	GnRH	GnRH-agonist		rHCG	r-HCG				GnRH-agonist	igonist	r-HCG		GnRH	GnRH-agonist		r-HCG	n-HCG	r-HCG	N/A	GnRH-agonist	N/A	N/A	N/A
Total oocytes retrieved	26	18	25	13	4	31	ŝ	14	29	21	S.	~	21	2	11	16	19	20	14	7 11	38	0	0
Total mature oocytes (MII)	17	12	23	12	-	30	m	11	25	16	4	~	10	4	œ	7	6	œ	00	4	N/A	0	0
OHSS	Ň	No	No	N/A	Ň	Yes	No	No	No	No	Ň		N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
<sup>a</sup> Not Available	a.																						
<sup>b</sup> The first cycle	ц																						
<sup>c</sup> Second cycle		-			:		1																

<sup>d</sup> Secondary pulmonary hypertension caused by transposition of great vessels

<sup>e</sup> pmol/L <sup>f</sup> Myelodysplastic/pre-malignant clone with monosomy 7

<sup>g</sup> nmol/L

the GnRH-antagonist protocol also provides the chance to utilize a GnRH-agonist for final oocyte maturation. Gonadotropin administration can begin at any point in the menstrual cycle. Response to medication will be evaluated with ultrasound (trans-vaginal or trans-abdominal, depending on the patient's comfort level) and E2 measurements, with gonadotropin dosage adjusted accordingly. For an OS protocol, once the leading follicle grew to at least 12 mm in diameter or E2 reached 300 pg/mL, the patient began a daily injection of GnRH-antagonist to prevent ovulation [11].

It is anticipated that many females with age under 18 years may have an immature HPO-axis [11]. There is no decisive data, whether FSH alone or FSH plus luteinizing hormone (LH) should be used in these cases. Due to the predicted immaturity of the HPO-axis, FSH plus LH administration is the preferred approach [16]. Also, one option to limit the risk of ovarian hyperstimulation syndrome (OHSS) is considered a GnRH-agonist trigger instead of HCG. However, there are some concerns to use GnRH-agonists in the adolescent population because of the HPO-axis immaturity [17]. Therefore, it is imperative to check serum progesterone and LH levels  $\sim 8{-}14$ h following a GnRH-agonist trigger to ensure that the triggering was effective [29, 30]. Typically an inadequate response is defined as a progesterone <3 ng/ml and an LH <15 IU/L [29].

Given the evidence so far equating a pregnancy rate of 6% per vitrified oocyte [31], around 20 oocytes being used per live birth [32]. The lower oocyte yields some oncologic patients deprived them of a good chance of pregnancy and live birth. However, in the poor response patients, there is the option of performing oocyte banking over a number of cycles [33]. Although, increasing the overall number of vitrified oocytes double stimulation during the follicular and luteal phases in the same cycle has been described successfully in these patients [34, 35].

Most studies on the OS outcomes and OC for adolescent girls with oncologic or non-oncologic diseases are limited to case reports [8, 13, 15–17] or case series [9, 10, 12, 14]. Based on the results of these studies, the hormonal profiles (FSH, LH, and AMH) and AFC of patients do not show significant differences in the adult patients. The OS protocol for patients have been OS with GnRH agonist or antagonist, which did not show a significant difference with adult patients in terms of duration of stimulation, gonadotropins dose, triggering medication, total oocytes retrieved, and total mature oocytes (MII).

Only Nagashima et al. reported no access to the oocyte in two patients (17 and 18 year old) with acute lymphoblastic leukemia (ALL) and acute myeloid leukemia (AML) [12]. Complications, such as OHSS,

are very rare. Only Lavery et al. reported a case of OHSS that needed treatment support [10]. No OHSS case was observed in our study.

Kim et al. reported a live birth from the frozen oocytes retrieved from a 17-year old adolescent with secondary pulmonary hypertension caused by transposition of great vessels. After 5 years, 11 oocytes were fertilized under microinjection process. Two 5-day embryos were transferred to the patient. The outcome was the birth of a healthy boy at 38 weeks of pregnancy [8].

Manuel et al. carried out the OS and OC on the 41 oncologic patients between the ages 13 and 21 years old. Among them, 38 patients underwent a successful procedure of retrieving and freezing mature oocytes.

When dividing patients to 13-17 yr. group and 18-21 yr. group, there was no statistical difference in the AMH level, peak E2, gonadotropin dosage, duration of stimulation, total oocytes retrieved, mature oocytes retrieve, and cry-opreservation between these two groups [11]. Hipp et al. compared the results of OS and OC in the patients under the age of 20 who were candidates for FP prior to gonado-toxic treatment with patients between the ages 20 and 29 years old. According to their report, even though the possibility of canceling the cycle of the OS due to the poor ovarian response is quite higher in the age group under 20, other factors of OS such as gonadotropins dosage, duration of OS, the number of retrieved oocytes, number of frozen mature oocytes are similar in both age groups [7].

In our study, the OS was performed in the five patients with HL. Comparison of these patients in terms of number of retrieved oocytes and mature oocytes suggests that the method of triggering final oocyte maturation is an effective factor in the number of mature oocytes.

In the present study, two patients had ovarian tumor (germ cell tumor and dysgerminoma) and a history of unilateral oophorectomy. Our oncologist consultation confirms the OS and vaginal oocyte retrieval in these patients. FP in ovarian cancer stands more challenging. In ovarian cancer patients, trans-vaginal oocyte retrieval carries a risk of ovarian capsule rupture and cancer cell spillage, which can cause staging up from 1A to 1C [36]. Bilateral disease is uncommon. The majority of patients have stage I disease, and nearly 90-95% of the cases are curable via post-operative chemotherapy) [37]. For the patient with germ cell tumor, the OS was carried out in the luteal phase and for the patient with dysgerminoma in the follicular phase. Ten oocytes were retrieved in the patient with germ cell tumor and 5 oocytes in the patient with dysgerminoma. Noyes et al. reported 38 retrieved oocytes in a 19-year-old patient with ovarian luteinized thecoma after OS with GnRH-agonist long protocol. Oktay et al. reported 8 retrieved oocytes (4 MII) in

a 15-year-old patient with germ cell tumor after OS with GnRH-antagonist protocol. Contrary to our study, they used human menopausal gonadotropin (HMG) and/or r-LH in addition to r-FSH for OS, and utilized GnRH-agonist for triggering final oocyte maturation. In our study, use of different triggering methods could be the reason for retrieving different numbers of oocytes from the two patients.

In this study, no oocytes were collected after OS of medulloblastoma patient. While, Kutteh et al. reported OS performance for three medulloblastoma patients and, a retrieved desirable number of oocytes. They used HMG for the OS, whereas r-FSH was utilized in our research. Considering the effects of a brain tumor on the hypothalamic-pituitary axis, it seems that combined the use of r-FSH and HMG/r-LH, and also use of HCG instead of GnRH-agonist for triggering, could lead to retrieval of oocyte in this patient. Considering the possibility of incomplete maturity of the HPO-axis in adolescents, and its compensation by simultaneous administration of gonadotropins containing FSH and LH, and also more focusing on the triggering method including use of a "dual triggering" method, can be accompanied by retrieval of a larger number of mature oocytes.

Some studies have proposed that the approach of OTC simultaneously with oocyte or embryo cryopreservation be used as a solution for increasing the efficiency of FP methods. For this aim, laparoscopic surgery is performed to remove half of the tissues of one ovary and OS begins after one to two days. Research has shown that this method is not accompanied so far by any known complications and the number of collected oocytes does not decrease considerably after removal of the ovarian tissues [38, 39]. Here, we used the same method for a patient who suffered of HL and also, 13 mature oocytes retrieved.

In our center, FP has not been done for adolescents who are FP candidates for non-cancer indication such as those with thalassemia major, sickle cell anemia, rheumatoid diseases, and Turner syndrome. As a result, the present study did not compare the OS results of these cancer adolescents with non-cancer peers.

Based on our result, we suggested that, this option of FP is a feasible procedure for this age group. Among the available studies, only Kim et al. reported the success on OC for an adolescent patient. Other studies, and present research, provided the report regarding the information about the OS cycle, such as prescribed gonadotropins dosage, OS duration, probable side effects of the OS, the number of retrieved and frozen matures oocytes, and EC in some cases.

The results of the available studies resolved the issues regarding the subjects such as lack of sufficient knowledge regarding the adolescent patients' response to OS, the possibility of failure of the HPOaxis to reach full maturity that might result in immature oocytes, plus, the uncertainty of the predictive factor of ovarian response in this age group including AMH and AFC. However, subjects such as uncertainty regarding the success rate of using the frozen oocyte from patients under the age of 18-year -old remain unanswered.

Based on the findings of the present study it is concluded that, OC is a feasible option in the adolescents. However, only if reported acceptable fertilization rates, as well as the successful cases of live birth from OC at the ages under 18, this option of preserving fertility can be applied to this age range. Therefore, we shall wait for more reports in the future regarding the use of frozen oocytes retrieved from patients under the age of 18-year-old, and then make the final judgment concerning the efficiency of this option for preserving fertility in this age range.

# Abbreviations

- FP Fertility preservation
- OTC Ovarian tissue cryopreservation
- EC Embryo cryopreservation
- OS Ovarian stimulation
- ASRM American Society for Reproductive Medicine
- OC Oocyte cryopreservation
- HPO Hypothalamus-pituitary-ovarian
- AMH Anti-mullerian hormone
- AFC Antral follicle count
- HL Hodgkin Lymphoma
- GnRH Gonadotropin releasing hormone
- r-FSH Recombinant follicle stimulating hormone
- r-HCG Recombinant human chorionic gonadotropin
- ASCO American Society of Clinical Oncology
- POF Premature ovarian failure
- LH Luteinizing hormone
- OHSS Ovarian hyperstimulation syndrome
- MII Mature oocytes
- ALL Acute lymphoblastic leukemia
- AML Acute myeloid leukemia
- HMG Human menopausal gonadotropin

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### Authors' contributions

FG: provided clinical expertise and supervision, project development, and manuscript editing; AY: project development, study design, data collection/ management and manuscript writing; MM: manuscript editing.

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No financial support has been granted.

# Availability of data and materials

All data generated or analyzed during this study are included in this published article [and its supplementary information files].

# Declarations

# Ethics approval and consent to participate

All he procedures performed in studies involving human participants were in accordance with the ethical standards committee of the Royan Institute and with the 1964 Helsinki Declaration and its later amendments or comparable ethical standards.

Ethics Approval Code: IR.ACECR.ROYAN.REC. 1400.080.

Informed consent was obtained from all the individual participants included in the study.

### **Consent for publication**

Written consent to publish this information was obtained from the parents of the study participants.

# **Competing interests**

The authors declare no competing interests.

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### References

- Rodriguez-Wallberg KA, Marklund A, Lundberg F, Wikander I, Milenkovic M, Anastacio A, et al. A prospective study of women and girls undergoing fertility preservation due to oncologic and non-oncologic indications in Sweden-Trends in patients' choices and benefit of the chosen methods after long-term follow up. Acta obstetricia et gynecologica Scandinavica. 2019;98(5):604–15. https://doi.org/10.1111/aogs.13559.
- Michalczyk K, Cymbaluk-Płoska A. Fertility Preservation and Long-Term Monitoring of Gonadotoxicity in Girls, Adolescents and Young Adults Undergoing Cancer Treatment. Cancers. 2021;13(2):202. https://doi.org/ 10.3390/cancers13020202.
- Mulder RL, Font-Gonzalez A, Hudson MM, Van Santen HM, Loeffen EA, Burns KC, et al. Fertility preservation for female patients with childhood, adolescent, and young adult cancer: recommendations from the PanCareLIFE consortium and the International Late Effects of Childhood Cancer Guideline Harmonization Group. Lancet Oncol. 2021;22(2):e45–56. https://doi.org/10.1016/S1470-2045(20)30594-5.
- Rodriguez-Wallberg KA, Anastacio A, Vonheim E, Deen S, Malmros J, Borgström B. Fertility preservation for young adults, adolescents, and children with cancer. Upsala J Med Sci. 2020;125(2):112–20. https://doi.org/10. 1080/03009734.2020.1737601.
- Medicine PCotASfR. Fertility preservation in patients undergoing gonadotoxic therapy or gonadectomy: a committee opinion. Fertil Steril. 2019;112(6):1022-33.https://doi.org/10.1016/j.fertnstert.2019.09.013.
- Rowell E, Duncan F, Laronda M. ASRM removes the experimental label from Ovarian Tissue Cryopreservation (OTC): pediatric research must continue. Fertil Steril Dialog. 2020. https://doi.org/10.1016/78659c74-03b7-4024-85ad-c628e2542107.
- Hipp HS, Shandley LM, Schirmer DA, McKenzie L, Kawwass JF. Oocyte cryopreservation in adolescent women. J Pediatr Adolesc Gynecol. 2019;32(4):377–82. https://doi.org/10.1016/j.jpag.2019.03.001.
- Kim TJ, Hong SW. Successful live birth from vitrified oocytes after 5 years of cryopreservation. J Assist Reprod Genet. 2011;28(1):73–6. https://doi. org/10.1007/s10815-010-9487-3.
- Kutteh WH, Klosky JL, Green DM, Sparrow CK, Kutteh MA, Robinson GW, et al. Ovulation induction and oocyte retrieval for fertility preservation in young adolescents newly diagnosed with medulloblastoma: a case series. J Obstet Gynaecol. 2018;38(6):878–9. https://doi.org/10.1080/ 01443615.2017.1398222.
- Lavery SA, Islam R, Hunt J, Carby A, Anderson RA. The medical and ethical challenges of fertility preservation in teenage girls: a case series of sickle cell anaemia patients prior to bone marrow transplant. Hum Reprod. 2016;31(7):1501–7. https://doi.org/10.1093/humrep/dew084.
- 11. Manuel SL, Moravek MB, Confino R, Smith KN, Lawson AK, Klock SC, et al. Ovarian stimulation is a safe and effective fertility preservation option

- Nagashima T, Muroi K, Kawano-Yamamoto C, Miyoshi T, Ohmine K, Toshima M, et al. Autologous gamete cryopreservation before hemopoietic stem cell transplantation. Med Sci Monit. 2005;11(3):CR91-CR4.
- Noyes N, Perretta RC, Fino ME, Matulewicz T, Barakat R. Use of hormone suppression then oocyte freezing to preserve reproductive capability in an adolescent girl with ovarian luteinized thecoma associated with sclerosing peritonitis. Fertil Steril. 2009;92(1):393. e11-. e14.https://doi.org/10. 1016/j.fertnstert.2009.01.145.
- Oktay K, Bedoschi G. Oocyte cryopreservation for fertility preservation in postpubertal female children at risk for premature ovarian failure due to accelerated follicle loss in Turner syndrome or cancer treatments. J Pediatr Adolesc Gynecol. 2014;27(6):342–6. https://doi.org/10.1016/j.jpag. 2014.01.003.
- Oktay K, Rodriguez-Wallberg KA, Sahin G. Fertility preservation by ovarian stimulation and oocyte cryopreservation in a 14-year-old adolescent with Turner syndrome mosaicism and impending premature ovarian failure. Fertility and sterility. 2010;94(2):753. e15-. e19.https://doi.org/10.1016/j. fertnstert.2010.01.044.
- Peddie VL, Maheshwari A. Successful controlled ovarian stimulation and vitrification of oocytes in an adolescent diagnosed with myelodysplastic/ pre-malignant clone with monosomy 7. Hum Fertil. 2018;21(1):39–44. https://doi.org/10.1080/14647273.2017.1347288.
- Reichman DE, Davis OK, Zaninovic N, Rosenwaks Z, Goldschlag DE. Fertility preservation using controlled ovarian hyperstimulation and oocyte cryopreservation in a premenarcheal female with myelodysplastic syndrome. Fertil Steril. 2012;98(5):1225–8. https://doi.org/10.1016/j.fertn stert.2012.07.1056.
- Coyne K, Purdy M, O'Leary K, Yaklic JL, Lindheim SR, Appiah LA. Challenges and considerations in optimizing ovarian stimulation protocols in oncofertility patients. Front Public Health. 2014;2:246.
- Stoop D, De Vos M, Tournaye H, Devroey P. Fertility preservation utilizing controlled ovarian hyperstimulation and oocyte cryopreservation in a premenarcheal female with myelodysplastic syndrome. Fertil Steril. 2012;98(5):1121–2. https://doi.org/10.1016/j.fertnstert.2012.07.1093.
- Ehrbar V, Urech C, Alder J, Harringer K, Dällenbach RZ, Rochlitz C, et al. Decision-making about fertility preservation—qualitative data on young cancer patients' attitudes and needs. Arch Women's Mental Health. 2016;19(4):695–9. https://doi.org/10.1007/s00737-016-0604-x.
- Oktay K, Harvey BE, Partridge AH, Quinn GP, Reinecke J, Taylor HS, et al. Fertility preservation in patients with cancer: ASCO clinical practice guideline update. J Clin Oncol. 2018;36(19):1994–2001. https://doi.org/10. 1200/JCO.2018.78.1914.
- Moravek MB, Appiah LC, Anazodo A, Burns KC, Gomez-Lobo V, Hoefgen HR, et al. Development of a pediatric fertility preservation program: a report from the pediatric initiative network of the oncofertility consortium. J Adolesc Health. 2019;64(5):563–73. https://doi.org/10.1016/j.jadoh ealth.2018.10.297.
- Ben-Aharon I, Abir R, Perl G, Stein J, Gilad G, Toledano H, et al. Optimizing the process of fertility preservation in pediatric female cancer patients–a multidisciplinary program. BMC Cancer. 2016;16(1):1–9. https://doi.org/ 10.1186/s12885-016-2584-7.
- Filippi F, Meazza C, Paffoni A, Raspagliesi F, Terenziani M, Somigliana E. Egg freezing in childhood and young adult cancer survivors. Pediatrics. 2016;138(4).https://doi.org/10.1542/peds.2016-0291.
- Friedler S, Koc O, Gidoni Y, Raziel A, Ron-El R. Ovarian response to stimulation for fertility preservation in women with malignant disease: a systematic review and meta-analysis. Fertil Steril. 2012;97(1):125–33. https://doi. org/10.1016/j.fertnstert.2011.10.014.
- Garcia-Velasco JA, Domingo J, Cobo A, Martínez M, Carmona L, Pellicer A. Five years' experience using oocyte vitrification to preserve fertility for medical and nonmedical indications. Fertil Steril. 2013;99(7):1994–9. https://doi.org/10.1016/j.fertnstert.2013.02.004.
- Quinn GP, Murphy D, Knapp C, Stearsman DK, Bradley-Klug KL, Sawczyn K, et al. Who decides? Decision making and fertility preservation in teens with cancer: a review of the literature. J Adolesc Health. 2011;49(4):337–46. https://doi.org/10.1016/j.jadohealth.2011.01.005.
- 28. Anderson RA, Wallace WHB. Antimüllerian hormone, the assessment of the ovarian reserve, and the reproductive outcome of the young patient

with cancer. Fertil Steril. 2013;99(6):1469–75. https://doi.org/10.1016/j. fertnstert.2013.03.014.

- Beall S, Moon K, Widra E, Segars J, Chang F, Richter K. Human chorionic gonadotropin (hCG) re-trigger following a poor response to leuprolide acetate (LA) trigger is not associated with poor in vitro fertilization (IVF) treatment outcomes. Fertil Steril. 2012;98(3):S52. https://doi.org/10. 1016/j.fertnstert.2012.07.187.
- Shapiro BS, Daneshmand ST, Restrepo H, Garner FC, Aguirre M, Hudson C. Efficacy of induced luteinizing hormone surge after "trigger" with gonadotropin-releasing hormone agonist. Fertil Steril. 2011;95(2):826–8. https://doi.org/10.1016/j.fertnstert.2010.09.009.
- Loren AW, Mangu PB, Beck LN, Brennan L, Magdalinski AJ, Partridge AH, et al. Fertility preservation for patients with cancer: American Society of Clinical Oncology clinical practice guideline update. J Clin Oncol. 2013;31(19):2500. https://doi.org/10.1200/JCO.2013.49.2678.
- Stoop D, Ermini B, Polyzos N, Haentjens P, De Vos M, Verheyen G, et al. Reproductive potential of a metaphase II oocyte retrieved after ovarian stimulation: an analysis of 23 354 ICSI cycles. Hum Reprod. 2012;27(7):2030–5. https://doi.org/10.1093/humrep/des131.
- Chung K, Donnez J, Ginsburg E, Meirow D. Emergency IVF versus ovarian tissue cryopreservation: decision making in fertility preservation for female cancer patients. Fertil steril. 2013;99(6):1534–42. https://doi.org/ 10.1016/j.fertnstert.2012.11.057.
- 34. Kuang Y, Chen Q, Hong Q, Lyu Q, Ai A, Fu Y, et al. Double stimulations during the follicular and luteal phases of poor responders in IVF/ICSI programmes (Shanghai protocol). Reprod Biomed Online. 2014;29(6):684–91. https://doi.org/10.1016/j.rbmo.2014.08.009.
- Moffat R, Pirtea P, Gayet V, Wolf JP, Chapron C, de Ziegler D. Dual ovarian stimulation is a new viable option for enhancing the oocyte yield when the time for assisted reproductive technnology is limited. Reprod Biomed Online. 2014;29(6):659–61. https://doi.org/10.1016/j.rbmo.2014.08.010.
- Kim S-Y, Lee JR. Fertility preservation option in young women with ovarian cancer. Future Oncol. 2016;12(14):1695–8. https://doi.org/10.2217/ fon-2016-0181.
- Billmire D, Vinocur C, Rescorla F, Cushing B, London W, Schlatter M, et al. Outcome and staging evaluation in malignant germ cell tumors of the ovary in children and adolescents: an intergroup study. J Pediatr Surg. 2004;39(3):424–9. https://doi.org/10.1016/j.jpedsurg.2003.11.027.
- Dolmans M-M, Marotta M-L, Pirard C, Donnez J, Donnez O. Ovarian tissue cryopreservation followed by controlled ovarian stimulation and pick-up of mature oocytes does not impair the number or quality of retrieved oocytes. J Ovarian Res. 2014;7(1):1–5. https://doi.org/10.1186/ s13048-014-0080-8.
- Huober-Zeeb C, Lawrenz B, Popovici RM, Strowitzki T, Germeyer A, Stute P, et al. Improving fertility preservation in cancer: ovarian tissue cryobanking followed by ovarian stimulation can be efficiently combined. Fertil Steril. 2011;95(1):342–4. https://doi.org/10.1016/j.fertnstert.2010.07.1074.

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