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Serum agonistic autoantibodies against type-1 angiotensin II receptor titer in patients with epithelial ovarian cancer: a potential role in tumor cell migration and angiogenesis

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Abstract

Background: Although agonistic autoantibodies against type-1 angiotensin-II receptor (AT₁-AA) are frequently detected in women with preeclampsia, the clinical significance of AT₁-AA in association with epithelial ovarian cancer (EOC) has not been identified.

Methods: In an attempt to clarify this issue, we measured serum AT₁-AA titer from EOC patients (n = 89) and healthy normal subjects (n = 55), correlated AT₁-AA titer with EOC stage and grade, and demonstrated the effects of purified AT₁-AA on migration of ovarian cancer cells and angiogenesis of chick embryo chorioallantoic membrane.

Results: We found that the AT₁-AA titer was significantly higher in EOC patients compared with healthy control subjects (1.77 ± 0.28 vs. 0.35 ± 0.05 , $P < 0.01$). The positive rate was averaged by $72.1 \pm 2.5\%$ in EOC patients and $15.5 \pm 1.5\%$ in control ($P < 0.01$). Increased AT₁-AA titer in EOC patients was associated with advanced stages and grades of EOC, and positively correlated with level of vascular endothelial growth factor ($r = 0.855$, $P < 0.01$). Furthermore, AT₁-AA directly stimulated migration of ovarian cancer cells and enhanced microvascular density of chick embryo chorioallantoic membrane. These AT₁-AA-mediated effects were significantly blocked either by an autoantibody-neutralizing peptide or an angiotensin II type I receptor antagonist, losartan.

Conclusion: Taken together, we found that a higher serum AT₁-AA titer may be associated with advanced progression of EOC in patients and play an important role in development of EOC by promoting cancer cell migration and angiogenesis. These findings implicate that AT₁-AA might be selected as a detectable biomarker and potential therapeutic target in diagnosis and treatment of EOC patients.

Keywords: Angiotensin II type I receptor, Autoantibodies against type-1 angiotensin II receptor, Angiogenesis, Epithelial ovarian cancer

Background

Epithelial ovarian cancer (EOC) is the sixth most common cancer and the seventh cause of death worldwide among women who develop gynecological cancer [1], with the estimated 22,280 new cases and 15,500 deaths in the United States in 2012. The vast majority of EOC patients are usually diagnosed with advanced stages due to the lack of adequate early screening tests and early specific symptoms during development of EOC [2]. The

standard treatment for advanced EOC patients includes debulking surgery followed by platinum–taxane based chemotherapy. These patients, however, are also at great risk of recurrence and emerging drug resistance with a more than 70% of relapse rate and a mean 18 months of progression-free survival period [2,3]. Therefore, understanding the pathogenesis of EOC and identifying early detectable biomarkers are essential to improve overall survival rate in advanced EOC patient.

It has well been defined that angiotensin II (Ang II) derived from the activated renin-angiotensin system plays a key role in the regulation of cardiovascular

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homeostasis through its two receptors: Ang II type 1 (AT₁) and type 2 (AT₂) receptors, which maintain arterial blood pressure, fluid and electrolyte homeostasis. Through the AT₂ receptor, Ang II evokes vasodilatation, sodium excretion and blood pressure reduction, and thereby counteracts the effects of AT₁ receptor [4]. However, increasing evidence suggests that Ang II is also involved in tumor cell migration/invasion, angiogenesis and metastasis through AT₁ receptor during the tumor development [5-7]. In patient with EOC, it has previously been reported that Ang II enhances vascular endothelial growth factor (VEGF) secretion, angiogenesis and tumor cell invasion via up-regulating G-protein-coupled AT₁ receptor; importantly, angiogenesis and peritoneal dissemination of the EOC can selectively be blocked using AT₁ receptor antagonist [6,8]. Therefore, considerable effort has been placed on the development of Ang II blockade therapy as a new strategy for EOC treatment.

Recent studies have demonstrated that agonistic autoantibodies against type-1 angiotensin II receptor (AT₁-AA) detected in preeclampsia induces significant placental trophoblast invasion [9], suggesting that AT₁-AA is one of the potential causative factors in development of preeclampsia. We have previously reported that AT₁-AA constricts human fetoplacental blood vessels and restricts fetal perfusion through activating Ang II AT₁ receptor [10]. Although animals studies have shown that activation of AT₁-AA is associated with elevation of intracellular Ca²⁺ in vascular smooth muscle cells [11], stimulation of placental and vascular NADPH oxidase [12] and activation of NF-κB [13], all of which may cause inflammation and contribute to pathogenesis of preeclampsia via AT₁-AA, there is less specific data to show whether AT₁-AA is elevated in patient with EOC and correlated with the advanced progression of EOC. Therefore, in the current study, we examined the serum AT₁-AA titer in EOC patients and determined whether change in AT₁-AA level is associated with malignant grades and angiogenic factor, VEGF. Using AT₁-AA purified from EOC patients, we demonstrated the effects of AT₁-AA on migration of ovarian cancer cells and microvascular density of chick embryo chorioallantoic membrane. Furthermore, we investigated whether the AT₁-AA-elicited biological effects could be suppressed by autoantibody-neutralizing AT₁-AA peptide, and whether cell migration and angiogenesis stimulated by AT₁-AA could be blocked by Ang II AT₁ receptor antagonist.

Methods

Patients

The study included 89 malignant EOC patients who were diagnosed and operated in the third hospital of Capital Medical University during the period of 05/2010 to 04/2012. Cases were chosen based on the histological grades and clinical stages of EOC patients according to

the International Federation of Gynecology and Obstetrics (FIGO) criteria. The healthy control subjects (n = 55) were enrolled from laparoscopy-negative cases on the clinical assessment at the same hospital. No significant difference in age was found between these two groups. The consent form was signed by all patients and the research protocol was approved by the Institutional Committee for the Protection of Human Subjects of Capital Medical University. Cases were excluded if patients were associated with 1) autoimmune diseases and endocrinal diseases; 2) complications derived from other different organ systems; 3) immune deficiencies diseases; 4) significant gastrointestinal diseases. All clinical and laboratory data were recorded. Serum samples were collected from the patients in both groups and stored at -80°C until use.

Measurement of AT₁-AA titer and VEGF by enzyme-linked immunosorbent assay (ELISA)

The serum AT₁-AA level in patients was detected by ELISA as we reported previously [10]. Briefly, 96-well microtiter plates were coated with 1 µg/ml AT₁R-ECII peptide synthesized from patients (GL Biochem Ltd, Shanghai, China) and incubated overnight at 4°C. After washing the plates with PBS three times, 50 µl serum samples were added to the plates and incubated at 37°C for 1 h. The biotinylated goat anti-human IgG antibody (1:3,000, Zhongshan Inc., Beijing, China) or streptavidin-peroxidase conjugate (1:2,000 Vector, CA, USA) was then incubated separately at 37°C for 1 h during washings. Finally, 2, 2'-azino-di (3-ethylbenzothiazoline) sulphonic acid (ABTS)-H₂O₂ (Roche, Basel, Switzerland) substrate buffer was applied for a half hour before reading. The optical densities (OD) from these plates were measured at 405 nm in a plate reader (Molecular Devices Corp, CA, USA). The AT₁-AA titer was expressed as the ratio of positive/negative (P/N), i.e., (the OD of specimen - the OD of blank control) / (the OD of negative control - the OD of blank control). The positivity of the serum sample to AT₁-AA was defined as P/N ≥ 2.1, while the negativity was defined as P/N ≤ 1.5. All assays were performed in duplicate. Commercially accessible ELISA kit (DaKeWe Biotechnological Corp, Shengzheng, China) were used to determine the patients' serum VEGF level according to the manufacturer's instructions. VEGF concentration was expressed as ng/L and the assays were performed in duplicate.

AT₁-AA peptide synthesis

AT₁-AA peptide fragments equivalent to the sequence of human anti-AT₁ receptor antibody (AT₁R-ECII, 165-191, I-H-R-N-V-F-F-I-I-N-T-N-I-T-V-C-A-F-H-Y-E-S-Q-N-S-T-L) was synthesized by solid-phase peptide synthesis method (GL Biochem Ltd, Shanghai, China). The purity of synthetic peptide was confirmed with a high pressure liquid chromatography as we reported previously [10].

Purification of the immunoglobulin G fraction

The total immunoglobulin G was isolated from serum samples with AT₁-AA positive EOC patients or AT₁-AA negative healthy normal subjects by Mab Trap Kit (Amersham, NJ, USA). The purities of extractions were assessed by sodium dodecylsulfonate–polyacrylate gel electrophoresis (SDS–PAGE) as we reported previously [10].

Cell lines and cell migration assay

Human ovarian cancer cells (OVCAR3) were purchased from the Cancer Hospital of Chinese Academy of Medical Sciences, Beijing, China and maintained in DMEM supplemented with 10% FBS, 2 mM l-glutamine, 100 units/ml penicillin and 100 µg/ml streptomycin. For all experiments, cells were detached with 0.25% trypsin and 0.02% EDTA and washed once in complete medium before use. Migration assay was conducted according to the manufacture's recommended protocol (BD Biosciences, New Jersey, USA). Briefly, OVCAR3 at 5×10^4 concentration were suspended in 300 µl of serum free media in the upper chamber with pre-coated filters (6.5 mm in diameter and 8 µm pore-size) with or without AT₁-AA, Ang II, AT₁R-ECII or Ang II AT₁ receptor antagonist, losartan. Bottom chambers were filled with medium containing 10% FBS as a chemoattractant. After cells were allowed to seed on the chambers for 24 h at 37°C, cells on the upper chamber and migrated cells at the bottom chamber were wiped with a cotton swab and then mixed with staining solution containing 0.125% coomassie blue in a mixture of methanol, acetic acid and water in a ratio of 45:10:45. The results were visualized under an inverted microscope from 5 randomized high power fields (x400). Results were calculated from the average of 3 separate assays conducted in triplicate.

Visualization of microvascular density in chick embryo chorioallantoic membrane (CAM)

Fertilized white leghorn chicken eggs were received at day 0 and incubated for 3 days at 37°C with constant humidity. On day 3, eggs were rinsed with 70% ethanol and a square window (0.5 cm²) was made with a pair of sterile scissor and cut away a circle of shell, thus exposing the underlying membrane (the chorioallantois). After the eggs (n = 8/each group) were treated with saline, AT₁-AA, Ang II, AT₁R-ECII or losartan, respectively for 30 min, the window was sealed with transparent tape and the eggs returned to the incubator at 90% relative humidity without turning. After 72 h of incubation, the CAM was fixed using 3.7% formaldehyde for 15 min, cut 3 cm² from the center and mounted on the slides for observation. The angiogenic results were visualized on an inverted microscope from 5 randomized fields. For each experiment, the staggered images were digitized and

results were calculated as a mean of microvascular density per high power field (x 400).

Statistical analysis

All data were calculated as mean ± SE. Statistical analysis was performed with SPSS 15.0 software. The positive rates in the two groups were compared with chi-square test. The *t*-test was applied for comparing two independent sample means, and the one-way ANOVA was used for comparing means of more than two samples. *P* < 0.05 was considered to be statistically significant.

Results

Clinical characteristics presented in EOC patients

Patient characteristics, stage and grade are shown in Table 1. The mean age of the EOC at primary diagnosis was 50.4±11 years and the mean history of the EOC was 7.4 ± 6 years. The mean age of the EOC at menarche was 15±2, and at menopause was 47±4. The FIGO stage of EOC patients was classified as follows: I: 6%; II: 56%; III: 23%; IV: 4%. Most EOC patients were at the grade III (61%). Fifty-four patients (28%) had ascites whereas 31% patients had no such complication.

Table 1 Patient characteristics

Characteristic	Number
Age at diagnosis	51 + 8.2
Weight (kg)	62 + 8.9
BMI (kg/m ²)	25 + 4.1
Age at menarche	15 + 2.0
Age at menopause	47 + 4.2
Stage	
I	6 (6.7%)
II	56 (62.9%)
III	23 (25.8%)
IV	4 (4.4%)
Grade	
G1	20 (22.4%)
G2	27 (30.3%)
G3	42 (85.7%)
Residual tumor	
≤2 cm	18 (20.2%)
>2 cm	71 (79.7%)
Ascites	
No	31 (34.8%)
Yes	58 (65.1%)
Diabetes Status	
No	30 (33.7%)
Yes	59 (66.2%)

Values are mean ± SE; others represent the percentage of total patients.

Clinical significance of AT₁-AA titer in EOC patients

The serum AT₁-AA titer in EOC patients and healthy control subjects was measured by ELISA. As shown in Figure 1A, the serum AT₁-AA titer was significantly increased from 0.35 ± 0.05 in healthy normal subjects to 1.77 ± 0.28 in EOC patients ($P < 0.01$). The average positive rate of AT₁-AA in EOC patients was significantly higher than that in healthy normal subjects ($72.1\% \pm 2.5\%$ vs. $15.5\% \pm 1.9\%$, $P < 0.01$). The correlation of serum AT₁-AA with clinicopathological outcomes was analyzed in EOC patients. As shown in Figure 1B, the number of AT₁-AA positive patients was increased with clinical FIGO stage: 45% in stage I, 61.5% in stage II and 72.8% in advanced stage III. Moreover, the AT₁-AA titer was also significantly higher in patients with an advanced grade (Figure 1C): 61.7% in grade I, 72.7% in grade II and 80.1% in grade 3. These results indicated that AT₁-AA level increases with progression of EOC stage and grade.

Correlation between serum AT₁-AA titer and VEGF

To determine whether serum AT₁-AA titer is associated with angiogenesis of the tumor, we examined the serum level of VEGF by ELISA in the same series of EOC patients. As shown in Figure 2A and 2B, VEGF level was significantly increased in patients with advanced FIGO stage and grade (i.e., at level III) compared with those in an early FIGO stage and grade (i.e., at level I). Positive linear correlation among the serum AT₁-AA level and VEGF was detected, (Figure 2C, $r^2 = 0.855$, $p < 0.01$), suggesting that AT₁-AA may play a role in angiogenesis during development of EOC through enhancing VEGF expression.

Effect of AT₁-AA on migration of ovarian cancer cells

OVCAR3 cells derived from the progressive adenocarcinoma of the ovary were used in this study. Migration of OVCAR3 cells stimulated by adding AT₁-AA was enhanced in a dose-dependent manner. As shown in the top panel of Figure 3, cell migration rates were consequently increased relative to the control when cells were treated with different dose of AT₁-AA (50, 100, 200 nM)

for 24 h. To demonstrate the potency of AT₁-AA in stimulation of cell migration by activating angiotensin AT₁ receptor, OVCAR3 cells were treated either with exogenous AT₁-AA (100 nM) or Ang II (100 nM), respectively before subjecting to cell migration. As shown at the bottom panel of Figure 3, both AT₁-AA and Ang II produced a comparable level in cell migration. Stimulation by AT₁-AA (100 nM) on cell migration was completely blocked either by the AT₁R-ECII (100 nM) or by the selective Ang II AT₁ receptor antagonist, losartan (300 nM), suggesting that AT₁-AA has direct stimulating effect on tumor cell migration and enhancement of OVCAR3 cell migration by AT₁-AA is mediated by Ang II AT₁ receptor. Addition of losartan or AT₁R-EC II alone had no effect on migration of OVCAR3 cells.

Effect of AT₁-AA on angiogenesis of the CAM

AT₁-AA administration caused a significant increase in microvascular density in the CAM. Figure 4 shows the representative photographs of AT₁-AA-treated and saline control CAM. Quantitatively, in each of the six experiments, the microvascular density of the CAM treated with AT₁-AA (100 nM) was increased by 60-70% compared with saline control. Addition of Ang II (100 nM) also increased the microvascular density of the CAM to a comparable level as that found in the AT₁-AA treated CAM. Enhancement in the microvascular density by AT₁-AA was significantly blocked either by AT₁R-ECII (100 nM) or losartan (300 nM), suggesting a role of AT₁-AA in angiogenesis through stimulating Ang II AT₁ receptor. Simultaneous addition of only the AT₁R-ECII (100 nM) or the losartan (300 nM), without AT₁-AA or Ang II, did not affect the microvascular density when compared with saline control (Figure 4).

Discussion

These results are the first to demonstrate that AT₁-AA level is significantly elevated in EOC patients. Enhanced AT₁-AA titer was associated with advanced stage and grade of the EOC and positively correlated with VEGF

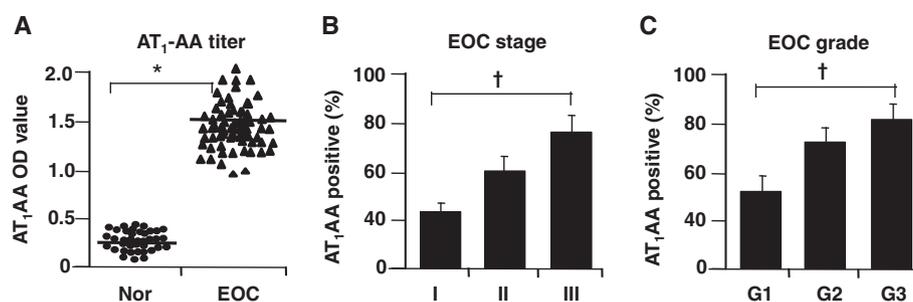


Figure 1 Level of AT₁-AA in EOC patients. Relative to the healthy normal subjects, AT₁-AA titer was significantly increased in EOC patients (A). The number of AT₁-AA positive patients was associated with clinical FIGO stage (B) and grade (C). Values are mean ± SE. * $p < 0.05$ vs. the normal subjects (Nor); † $P < 0.05$ vs. the stage I and grade I, respectively.

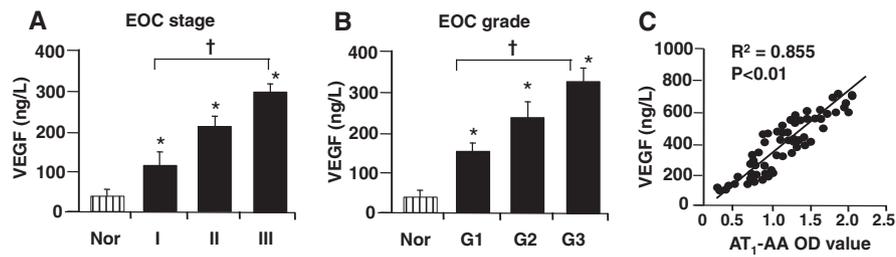


Figure 2 VEGF level in EOC patients and healthy normal subjects. Increased level of VEGF was detected in advanced stage (A) and grade (B). Scatter plots showed a positive linear correlation between VEGF level and AT₁-AA titer in EOC patients (C). Values are mean ± SE. *p < 0.05 vs. the normal subjects (Nor); † P < 0.05 vs. the stage I and grade I, respectively.

level in patients. Using cultured OVCAR3 cells and the CAM of chick embryo, we found that AT₁-AA has direct effect on cell migration and angiogenesis through activating Ang II AT₁ receptor.

AT₁-AA, an autoantibody against angiotensin II type 1 receptor, which is characterized to activate the receptor via specifically interacting with the second extracellular loop of the Ang II AT₁ receptor, has been documented to play a role in the pathogenesis of preeclampsia and hypertension [10,11,13-15]. However, AT₁-AA level and function has not been examined or identified in the ovarian cancer. In the current study, we found that serum titer and positive rate of AT₁-AA were significantly increased

in EOC patients. More importantly, this study revealed that the level of AT₁-AA is significantly elevated with an advanced FIGO stage and grade in EOC patients, supporting the concept that AT₁-AA may participate in ovarian cancer development and progression. As it has well been demonstrated, the FIGO stage and grade are poor prognostic factors for overall survival in EOC patients [3]. Therefore, monitoring serum AT₁-AA level might be of great value as a single marker in detecting all stages of EOC patients for clinical screening test, diagnosis and prognosis after therapeutic intervention.

VEGF is a main angiogenic factor in development of ovarian cancer through promoting angiogenesis and

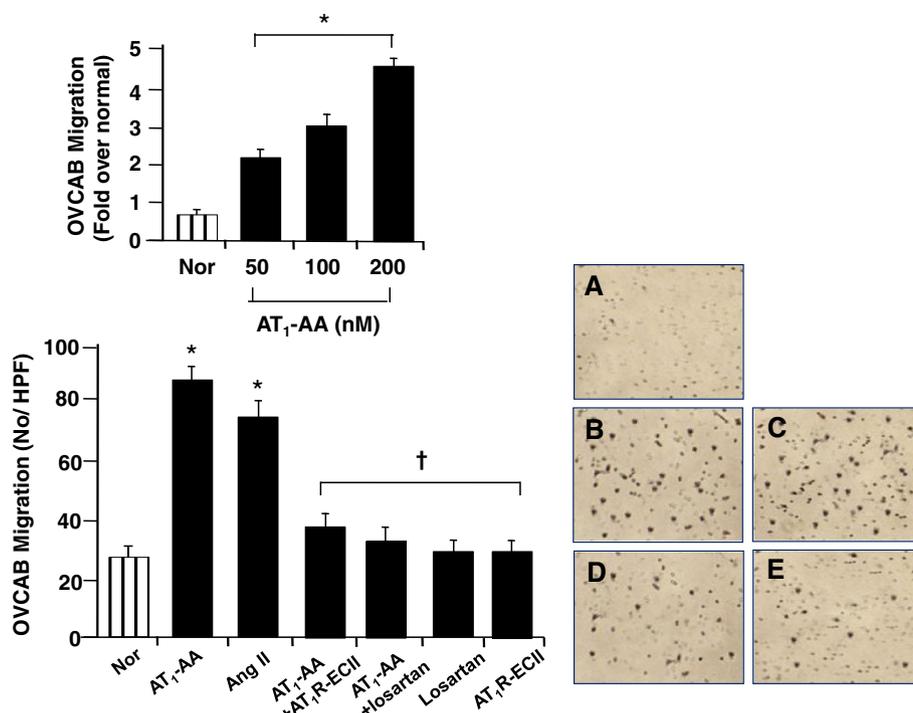


Figure 3 Effect of AT₁-AA on migration of OVCAR3 cells. AT₁-AA increased migration in a dose-dependent manner (top panel). Relative to the control (A), cell migration by AT₁-AA was significantly enhanced (B), which is comparable to the level found in Ang II (C). Enhancement of migration by AT₁-AA was blocked either by AT₁-AA-ECII (D) or losartan (E). Values are mean ± SE. n = 3 for each group. *p < 0.05 vs. saline control (Nor); †P < 0.05 vs. AT₁-AA and Ang II. HPF = high power field (x 400).

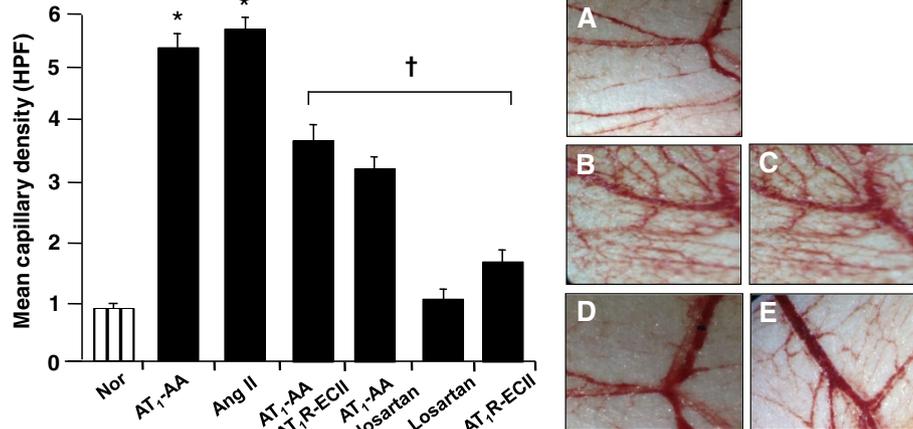


Figure 4 Effect of AT₁-AA on angiogenesis of the CAM. Relative to the saline control (A), AT₁-AA significantly increased microvascular density of the CAM (B) to a comparable level as did by Ang II (C). Enhancement of microvascular density by AT₁-AA was blocked either by AT₁-AA-ECII (D) or Ang II AT₁ receptor antagonist, losartan (E). Values are mean ± SE. n = 6 for each group. *p < 0.05 vs. saline control (Nor); †p < 0.05 vs. AT₁-AA and Ang II. HPF = high power field (x400).

significantly associated with tumor progression and poor prognosis [16-18]. Recent studies have shown that targeting inhibition of tumor angiogenesis through VEGF and its various signaling pathways is an effective therapy to suppress tumor growth and progression [8,17]. Our results showed that higher AT₁-AA titer is positively correlated with VEGF level in advanced stages of EOC patients, consistent with previous findings showing a role of Ang II in cancer development through VEGF gene expression and secretion [8,17].

Stimulation of AT₁ receptor by Ang II has been reported to be involved in tumor progression in a number of cancers including EOC [12,13]. The postulated role of AT₁-AA in cell migration and tumor spread led us to test if AT₁-AA has direct stimulating effect on ovarian cell migration. We selected either autoantibody-neutralizing AT₁-AA peptide, AT₁R-ECII as an inhibitor or selective AT₁ receptor antagonist, losartan to test the direct effect of AT₁-AA on cell migration and illustrate if this process is mediated by AT₁ receptor [11]. We found that the migratory number of OVCAR3 cells was significantly increased in AT₁-AA treated group, which was blocked either by AT₁R-ECII or losartan. These data suggested that AT₁-AA has direct effect on migration of ovarian cancer cells through activating AT₁ receptor, consistent with a previous report showing that Ang II-induced tumor cell invasion, angiogenesis and peritoneal dissemination are blocked by Ang II AT₁-receptor antagonist [19]. However, mechanistic studies are needed to further elucidate how AT₁-AA activates the Ang II AT₁ receptor. In line with our data, it has previously postulated that AT₁-AA may alter the structural conformation of Ang II AT₁ receptor so that the receptor's ability binding to circulating Ang II is enhanced [12].

The CAM of chick embryo has widely been selected to study the morphological aspects of tumor angiogenesis and metastasis [20]. We chose the CAM of chick embryo as a test model to demonstrate angiogenic substances in our study because of its extensive vascularization and easy accessibility to investigate mechanisms of action of proangiogenic and antiangiogenic molecules [20]. We found that addition of AT₁-AA at the same dose that causes OVCAR3 cell migration is effective in stimulating angiogenesis in the CAM, which was parallel with data showing elevation of VEGF in EOC patients. This increased microvascular density elicited by AT₁-AA was comparable to the level as that in the Ang II group. Furthermore, we showed that the use of AT₁R-ECII or AT₁ receptor blocker, losartan completely inhibits AT₁-AA-induced angiogenesis of the CAM. These findings suggest that an enhancement of angiogenesis by AT₁-AA involves activation of Ang II AT₁ receptor, thus selective Ang II AT₁ blockade therapy could efficiently inhibit the AT₁-AA-elicited angiogenesis under conditions exposed to AT₁-AA as it has previously been reported [19].

There are several limitations to this study that need to be mentioned. First, although *in vitro* studies speculated the mechanisms responsible for the migration of cancer cells and angiogenesis through AT₁ receptor, this study did not measure AT₁ receptor expression to show whether such a change is associated with AT₁-AA-mediated effects. Second, although a raised titer of AT₁-AA was detected in EOC patients, the "cause-effect" relationship remains to be investigated. In this regard, it will be interesting to determine whether the AT₁-AA titer falls in patients undergoing treatment. Third, the size of the study population was relatively small and limited only in the Asian patients. Therefore, future large-scale

clinical trials will be necessary to further determine whether AT₁-AA titer is also altered in EOC patients of different ethnicities.

Conclusions

In summary, we found that serum AT₁-AA is elevated in higher proportion of EOC patients, which is associated with advanced stages and pathological grades of EOC, and appears to promote the ovarian cell migration and angiogenesis through Ang II AT₁ receptor. This study provides promising data showing that AT₁-AA may play a significant role in development and progression of EOC, and might be considered as a potential therapeutic target in treatment of EOC patients.

Abbreviations

AT₁-AA: Agonistic autoantibodies against type-1 angiotensin-II receptor; Ang II: Angiotensin II; AT₁: Angiotensin II type 1 receptor; AT₂: Angiotensin II type II receptor; CAM: Chick embryo chorioallantoic membrane; EOC: Epithelial ovarian cancer; FIGO: International Federation of Gynecology and Obstetrics; OVCAR3: Human ovarian cancer cells; VEGF: Vascular endothelial growth factor.

Competing interests

The authors declare that there is no conflict of interest that would prejudice the impartiality of this research work.

Authors' contributions

LS and HRL participated in research design, patient's investigation and manuscript writing. JY and SLZ carried out the in vitro experiments and data acquisition; HYY and TL performed data analysis and interpretation. All authors read and approved the final manuscript.

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