

REVIEW

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The Jun N-terminal kinases signaling pathway plays a “seesaw” role in ovarian carcinoma: a molecular aspect

Yingyu Dou¹, Xiaoyan Jiang¹, Hui Xie¹, Junyu He² and Songshu Xiao^{1*}

Abstract

Ovarian cancer is the most common gynecological malignancy that causes cancer-related deaths in women today; this being the case, developing an understanding of ovarian cancer has become one of the major driving forces behind cancer research overall. Moreover, such research over the last 20 years has shown that the Jun N-terminal kinase (JNK) signaling pathway plays an important role in regulating cell death, survival, growth and proliferation in the mitogen-activated protein kinases (MAPK) signaling pathway, an important pathway in the formation of cancer. Furthermore, the JNK signaling pathway is often regulated by an abnormal activation in human tumors and is frequently reported in the literature for its effect on the progression of ovarian cancer. Although the FDA has approved some JNK inhibitors for melanoma, the agency has not approved JNK inhibitors for ovarian cancer. However, there are some experimental data on inhibitors and activators of the JNK signaling pathway in ovarian cancer, but related clinical trials need to be further improved. Although the Jun N-terminal kinase (JNK) signaling pathway is implicated in the formation of cancer in general, research has also indicated that it has a role in suppressing cancer as well. Here, we summarize this seemingly contradictory role of the JNK signaling pathway in ovarian cancer, that ‘seesaws’ between promoting and suppressing cancer, as well as summarizing the application of several JNK pathway inhibitors in cancer in general, and ovarian cancer in particular.

Keywords: Jun N-terminal kinases pathway, Ovarian cancer, Seesaw role, Anticancer effect, Tumor-promoting effect

Highlights

The JNK signaling pathway is abnormally activated in patients with ovarian cancer or drug-resistant ovarian cancer.

Autophagy mediated by the JNK signaling pathway plays a dual role in ovarian cancer.

The timing of influencing the JNK signaling pathway will affect the follow-up therapeutic effect.

Introduction

Ovarian carcinoma (OC) is one of the most common of the gynecologic cancers as well as being the most prevalent cause of gynecology tumor-related deaths worldwide [1]. To date there are some 239,000 new cases and 152,000 deaths due to OC each year [2]. In the United States

during 2018 there were about 22,240 new OC cases resulting in 14,070 deaths [3]. Whilst in Europe [1], the OC incidence rate is from 6.0 to 11.4 per 100,000 women, and although it is relatively lower in China, there was at least [4] 52,100 new cases and 22,500 deaths in 2015 alone. Most ovarian carcinomas are diagnosed at an advanced stage, of which 51% are diagnosed at stage III and 29% are diagnosed at stage IV [3, 5] and what are the risk factors for such incidence levels of OC?

Age growth, overweight or obesity, first full-term pregnancy after age 35, fertility therapy, hormone therapy after menopause, family history of OC, breast cancer or colorectal cancer might all be high risk factors for OC [6]. In addition, about 50% of OC patients are more than 65 years old [7] and according to early studies in the Netherlands, patients with stage II and III ovarian cancer, even in the absence of comorbidities, did not achieve the same effective as younger patients [8]. This difference may be related to the relatively poorer

* Correspondence: xiaosquirrel@163.com

¹Department of Gynecology and Obstetrics, the third Xiangya Hospital, the Central South University, Changsha 410013, Hunan, China
Full list of author information is available at the end of the article



physical conditions of the elderly [8]. However, the latest study indicates that older women with OC are 50% less likely to receive standard treatment than younger women, regardless of the type of treatment. Furthermore, when elderly patients receive personalized treatment, it has been shown that the treatment effect on them can be significantly improved [9, 10]. Age itself may not be a high-risk factor [11] and the etiology of OC is unclear but 5–10% of OC is thought to be hereditary. Hereditary OC, like breast cancer, is an autosomal dominant inheritance due to mutations in the BRCA1 and BRCA2 genes. Such gene mutations change the biological effects of cell tissues and, thus, play an indispensable role in promoting the occurrence and development of tumors. According to the dualism of OC, it can be divided into type I ovarian cancer and type II ovarian cancer. Concerning type I OC, the main gene mutations are KRAS, BRAF, PTEN, ARID1A, and PIK3CA, and its onset is slow, the diagnosis is mostly in the early clinical stage, and the prognosis is good. The main mutations in type II OC, however, are TP53 and BRCA1/2 and the onset of the disease is fast, aggressive, no prodromal symptoms, the diagnosis is mostly in the late clinical stage. Ovarian tissue composition is very complex, and it is the organ with the most types of primary tumors of all the organs of the body. There are great differences in different types of histological structure and biological behavior. According to the histological classification of the World Health Organization (WHO) 2014 edition, ovarian tumors can be divided into 14 categories, the main histological types of which are epithelial tumors, germ cell tumors and cord-stromal tumors. Epithelial tumors are the most common histological type of ovarian tumors, and their histology can be further divided into serous, mucinous and endometrioid types. Serous tumors are the main type of ovarian cancer. In addition, the five-year survival rate of serous cancer is 43%, while that of other types of endometrioid cancer is 82%, and that of mucinous cancer is 71% [3].

Although the optimal treatment is surgery as the main component, supplemented with appropriate chemotherapy such as TC (Paclitaxel and carboplatin), TP (Paclitaxel and cisplatin) and PC (cisplatin and cyclophosphamide), in about 70% of patients the ovarian cancer will reoccur. Clinical trials are trying new therapies and drug combinations, such as immunotherapy, target therapy, PARP and anti-angiogenesis drugs. At present, many experiments and clinical trials have been carried out to improve the therapeutic effect on ovarian cancer with the single or combined treatment of potential candidates, and some results have been achieved. However, much remains to be done. Specifically, understanding the genes of abnormally activated or expressed signal transduction pathways in the development of ovarian cancer will be helpful to improve

the prognosis of ovarian cancer and the development of new therapies. In this regard, and according to the latest cancer genome research, many signaling pathways have malfunctioned in the development of cancer [12, 13]. For instance, the abnormal activation of the Jun N-terminal kinases signaling pathway, one of the MAPK signaling pathways, has been reported in ovarian cancer frequently, making it one of the most important signaling pathways in the treatment of ovarian cancer [14–17]. To inform our discussions in this paper on the roles of JNK in the death of cells, cancer cell growth, and in chemotherapy resistance, it is worthwhile to first summarize the salient features of both the MAPK and JNK signaling pathways.

Key features of the MAPK and the JNK signaling pathway

The MAPK pathway includes a series of protein kinases such as ERK1/2, p38 $\alpha/\beta/\gamma/\delta$ MAPK and c-Jun amino terminal kinase 1/2/3 (JNK1/2/3), which control cell proliferation, cell survival and cell death in various processes [18, 19]. JNKs, also called ‘stress activated’ protein kinases just like p38 MAPKs, responds to various extracellular stimuli in different organisms.

JNK1/2/3 have been demonstrated in mammals, where splices have revealed more than ten various subtypes of transcription. And the proteins size of JNKs range from fourty-six kDa to fifty-five kDa. JNKs can be activated by cascade phosphorylation reaction of MAPKKK and MAPKK. More than 20 types of MAPKKK have been found such as the MEKK family members protein and the apoptosis related kinases pathway, which regulates and phosphorylates two different types of MAPKK, MKK4/7. MKK4 and MKK7, and activates JNK via phosphorylating the motifs of conserved tripeptide, that is tyrosine 185 and threonine 183 residues [19, 20]. Moreover, scaffold protein, nuclear factor-kB and dual specificity phosphatases also have an effect on the activity of JNKs [19]. Various constitutions of MAPKKK and MAPKK can control JNK signaling. Apoptosis signal regulation kinase 1 (ASK1) is in an inactive state in the normal conditions via interacting with thioredoxin [21]. ASK1/MAPKKK5 activating JNK is the mechanism of some kidney diseases [22]. Cypermethrin impairs astrocytes and disrupts the development of the extracellular-matrix via regulating reactive oxygen species (ROS), Ca²⁺ and JNK pathway [23]. Continuous activation of JNK in hepatocytes can lead to cell death or metabolic abnormalities [24]. TNF related apoptosis-inducing-ligand promotes apoptosis of Bim, homologue of Bcl-2, by activating JNK and its downstream substrates, and enhances hepatocyte cell anti-Fas-induced apoptosis [25]. Increasing ROS inhibits the phosphatase of JNK such as mitogen-activated protein kinase phosphatase 1 (MKP1 /DUSP1), which contributes to the continuous activation

of JNK [26]. ROS can also remove the inhibition of ASK1 [27]. Furthermore, MAP 2 K/MAP 3 K, interleukin-1, epidermal growth factor, drugs, endoplasmic reticulum stress (ER stress) and environment stress such as heat, hypoxia as well as radiation can also activate JNK.

There are so many molecular such as STAT1/3, c-jun, c-Myc, FOXO4, Bcl-2, ATF2, Smad2/3, PPAR γ 1 and RXR α that have been demonstrated as the JNK substrates. JNKs which increase the activity of c-jun transcription via binding and phosphorylating c-jun at Ser73 and Ser63 [28]. C-jun, a part of AP-1 that regulates expression of gene. AP-1, like JNKs can be activated by radiation, various stress or inflammatory cytokine. The activity of c-jun is essential for the Ha-Ras mediating carcinogenesis transformation [29]. Phosphorylated c-jun at Ser73 and Ser63 via Ha-Ras, c-Raf and v-Src induced carcinoma transformation in embryonic fibroblasts [30, 31]. And JNK regulates the activating-transcription factor 2 (ATF-2) and intracellular mitochondrial signaling pathway such as target proteins Sab [32, 33]. Using inhibitory peptide of Tat-Sab interferes with the activity of JNK blocking the increase of ROS.

However, some evidence also shows contradictory phenomena that lacking the JNK1/2 gene induces fetal death in mice. JNKs also control cell autophagy, proliferation or migration. Furthermore, experiments involving the editing of mouse genes suggest that overactivation or loss of the JNK pathway function induces the growth of cancer, inflammation and metabolism related diseases. Some evidence supports the idea that JNK has the function of promoting tumor development, whilst other evidence demonstrate that JNK plays an important role in suppressing cancer. Note that, c-jun is highly expressed in ovarian cancer tissues [34]. Furthermore, the JNK signaling pathway is abnormally active in platinum-resistant ovarian cancer, but platinum drugs can also promote apoptosis of ovarian cancer cells by activating the JNK signaling pathway [35–37]. Because the effect of the JNK signaling pathway in carcinoma is extremely complex but crucial, it may, therefore, be a potential target for molecular cancer treatment therapy.

JNK and ovarian tumor: role in the death

Cell death is important for maintaining the intracellular homeostasis environment and is critically regulated by the signaling pathway. The ability to increase the number of tumor cells depends not only on the rate of cell proliferation, but also on the rate of cell depletion [38]. Once the signal-regulated pathways of apoptosis, autophagy and other cell functions become abnormal, this may weaken the cell depletion rate for tumorigenesis so as to accelerate the development of cancer. Cell death includes necrosis and apoptosis. Cell apoptosis, a programmed death, is characterized by changes in the nucleus and forming

apoptosis body. The death receptor pathway, mitochondrial pathway and endoplasmic reticulum pathway can cause cell death, in addition, autophagy can also cause cell apoptosis.

After the discovery of lysosomes, the Greek term autophagy was proposed. Autophagy can be divided into auto and phagy according to the root, which refers to the process in which organelles in the cytoplasm are transported to lysosome and degraded [39]. Autophagy is formed by double membrane vesicles and lysosomes, namely the autophagy body, which swallows cell damaged or harmful protein, organelles and complexes and the damaged organelles and protein is degraded by autophagy body. Although autophagy protects both normal and cancer cells from apoptosis, especially in the environment of cytotoxicity, malnutrition and other stimuli, when exceeding a certain limit, autophagy can cause apoptosis of normal and cancer cells [40–42]. Autophagy can generally be divided into at least three major categories: macrophage, micro-autophagy and chaperone-mediated autophagy [43]. Usually autophagy refers to macrophage. Many researches have reported that autophagy plays a dual role, helping in cell survival or inducing cell death, depending on the strength or kinds of stimuli and the type of cell.

The JNK pathway plays an essential role in OC autophagy and cell death. Autophagy mediated by the JNK signaling pathway, not only can help OC cells avoid apoptosis when the tumor cells are in a nutrient-poor or cytotoxicity environment, but also can induce autophagy-mediated cell death. Activating JNK signaling pathway induce cell death of autophagy in OC cell, which through promoting conversion of LC3-I to LC3-II as well as forming autophagosomes, accompanying with inhibition of cell cycle in G1 stage via up-regulating the expression of p27 and p21 [44]. Unfolded protein response (UPR) and inositol-requiring enzyme 1 α (IRE1 α) are essential to autophagy. The character of the endoplasmic reticulum stress (ER stress) response, caused by nutrient deprivation, hypoxia, mitochondrial dysfunction, chemotherapy and etc., can enable the activation of PERK (double-stranded RNA activated protein kinase-like ER kinase), IRE1 α and ATF6 and is also controlled by JNK [42]. ER stress results in the accumulation of incorrect folded proteins in cell, although there are some ways help deal with those proteins such as UPR mediated by three different ER transmembrane receptors, ATF, PERK and IRE1, when these protective responses are not enough to cope with ER stress, cells would go through apoptosis in the end [45]. IRE1 α is also an essential regulator of the JNK pathway. UPR autophagy-mediated cell death might rely on influencing the activity of IRE1 α -JNK [46]. The chaperone proteins Gadd153 and GRP78 of ER also play an important role in UPR-induced cell death or survival [46, 47]. When

UPR in cells exceed a certain limit, damaged cells will die. This may be due to the activation of JNK/AP-1/Gadd153 so as to inhibit the expression of NF-kappa B or Bcl-2 and mediation of ATF6 and ATF4 in OC cells [46, 48]. Furthermore, it is reported that ER stress could not induce apoptosis in caspase-12 deficient cells [49]. This result might be due to IRE1 needing to gather TRAF2 as well as caspase-12 so as to induce cell apoptosis [49, 50]. The use of drugs related to ER stress or autophagy and silencing of autophagy-related proteins Beclin 1 and LC3 can increase cell survival in OC cells [42].

Low-glucose not only induces cell apoptosis via ER stress /IRE1 α /JNK, but also influences the generation of cell ATP resulting in cell death via the activation of ASK1 [51]. ASK1 activation leads to sustaining the activation of JNK [51, 52]. And energy metabolism reprogramming is one of the hallmarks of cancer [38]. Dr. Otto Warburg pointed out that cancer cells mainly rely on glycolysis. Furthermore, due to the low efficiency of glycolysis, a high rate for taking up glucose is essential for cancer cells [53]. Low glucose environments could, therefore, enhance some drugs cytotoxicity such as metformin via the energetic stress producing reactive oxygen species (ROS), influencing the mitochondria membrane potential and other byproducts of damaged cells originating from the process of mitochondrial oxidative phosphorylation [51, 54]. ROS-mediated cell apoptosis might, therefore, activate the ASK1/MKK4/7/JNK/mitochondrial signaling pathway [36]. AIF induces apoptosis through the mitochondrial pathway from mitochondrial translocation to the nucleus [55]. Furthermore, Bcl-2 expression decreases, Bax/Bak expression increases, while caspase 3/8/9 does not significantly change [56–58]. SP600125, a specific inhibitor of JNK/SAPK, prevents the expression of Bcl-2, increases the expression of Bax and AIF nuclear translocation, so as to reduce cell apoptosis [59]. Activation transcription factor 2(ATF-2), a component of AP-1, activates the JNK/SAPK pathway-mediated apoptosis of cells via increasing the trimethylation of histone H3K9 associated with the AP-1 binding region interacting with the Bcl-2 promoter [58]. Inhibition of the Akt signaling pathway and c-FLIPL activation can induce the apoptosis of OC cells through up-regulation of p-JNK and subsequent activation of caspase-3 [60]. However, there are also conflicting experimental results suggesting that inhibiting JNK activity may promote cancer cell death. SP600458 can significantly enhance the death of OC cells caused by PARP1 protein cleavage and caspase-3 by disrupting MMP in nilotinib [61]. This may be due to the different subtypes of JNK that play a role. It has been pointed out that WBZ_4, a new inhibitor of JNK1, or targeting A2780CP20, SKOV3 and HEYA8 with JNK1 siRNA, can significantly inhibit the proliferation of OC cells [62].

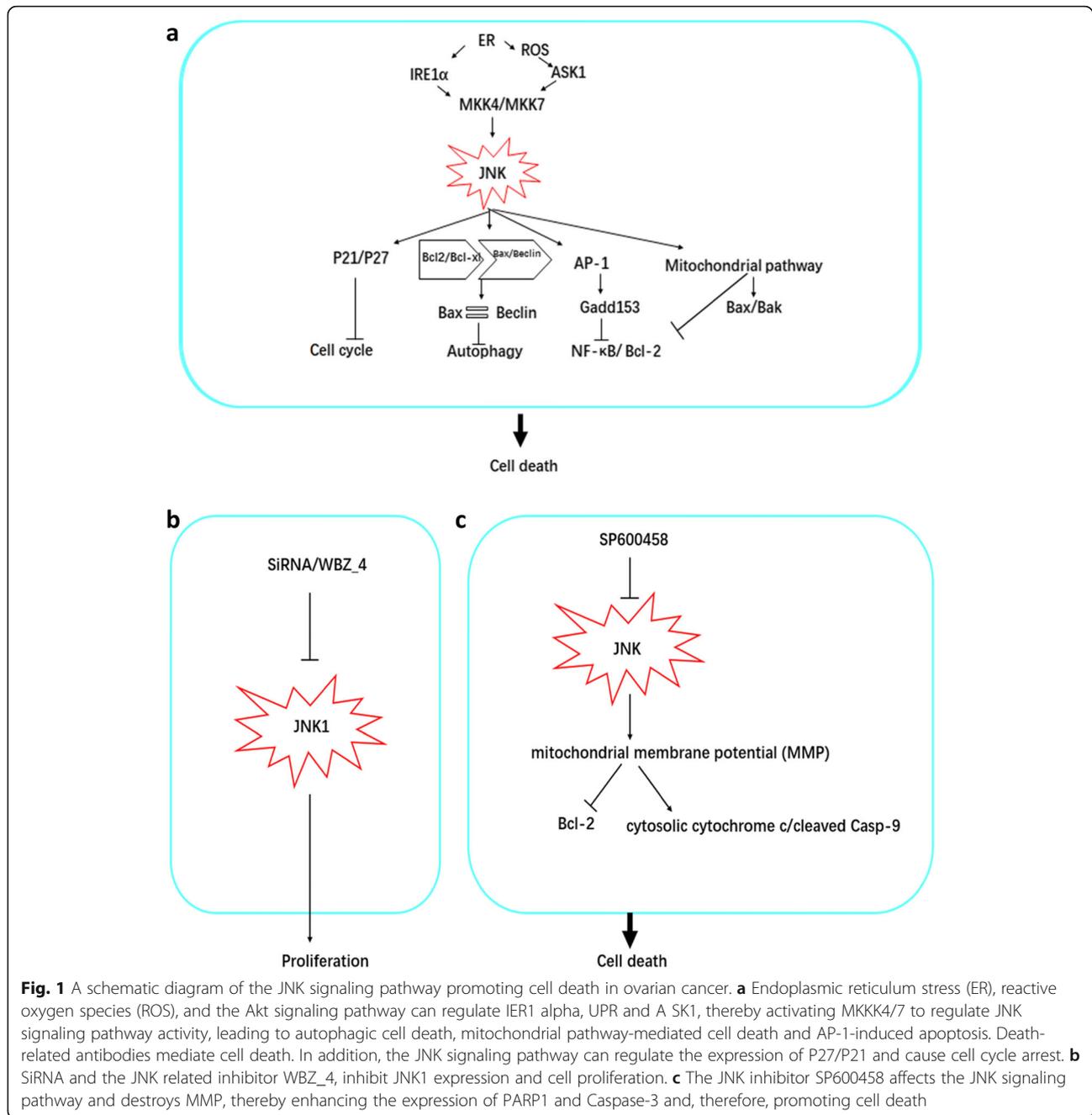
Furthermore, a clinical trial of ovarian cancer (NCT01015118) indicated that, Nintedanib, which is a triple vascular kinase inhibitor of the VEGF receptor, a platelet-derived growth factor receptor and a fibroblast growth factor receptor, combined with carboplatin and paclitaxel can significantly improve progression-free survival in patients with advanced ovarian cancer [63, 64]. Nintedanib can up-regulate the expression of pulmonary surfactant protein D in A549 cells through the JNK/AP-1 pathway, and alleviate lung fibrosis, however, this does not affect cell proliferation [65] but it can inhibit the proliferation of prostate cancer cells [66]. Concerning ovarian cancer, it is clear that further research is needed to demonstrate how Nintedanib affects the development of ovarian cancer through JNK signaling pathway.

At present, there are only a few reports on JNK inhibitors in the treatment of ovarian cancer; this may be due to the dual role of the JNK signaling pathway, discussed above, in promoting ovarian cancer apoptosis. For instance, mice lacking only one mutant of *jnk1/2/3* and either a mutant of *jnk1/jnk3* or *jnk2/jnk3* can survive normally, but lacking JNK1 and *jnk2* at the same time leads to the death of mice embryos and a very serious abnormal apoptosis in their brain cells [67]. In addition, the phosphorylation of JNK1, JNK2 and c-jun is necessary for ultraviolet-induced apoptosis, but JNK2 activated by ER stress can promote the survival of cells [36, 68]. The mechanism of JNK-induced cell death in ovarian cancer is depicted in Fig. 1.

The function of JNK in the regulation ovarian cancer cell growth

Normal cells need to be regulated by growth signals to change from a stationary state to a proliferative state. Without the stimulation of such signals, no normal cells will proliferate [69]. However, for cancer cells, the proliferation behavior relying on endogenous growth signals is greatly reduced, probably because cancer cells mainly rely on the regulation of carcinogenic genes and their own growth signals. In addition to self-sufficient growth signals and gene mutations, autophagy may also promote tumorigenesis and development by avoiding cell death through internal regulation.

The type of high level serous ovarian cancer with highly aggressive ability is the cause of most, 70–80% [70], of ovarian cancer deaths. Furthermore, this pathological type is the most common type of ovarian cancer. TP53 mutation may induce cell loss through the normal function of anti-cancer, which covers or masks the role of the p53 in normal cell. Carcinomas with TP53 mutations generally have the characteristics of high aggressiveness, invasiveness, and poor-differentiation. According to the report, the frequency of mutation in ovarian cancer ranges from about 50 to 100% [70, 71], such a high rate is



attributed to the different histological types of ovarian cancers having different frequencies of mutation. It shows that the frequency of TP53 mutation is rare in low level serous cancer or borderline carcinomas, whereas TP53 mutation is very common in high level serous ovarian cancer, even reaching 100% in some samples [71, 72]. The mutation of TP53 can weaken JNK/ROS/TP53 signaling pathway-mediating cell apoptosis. The tumorigenic effect of JNK is not only weakened by inhibition activity, but also can be promoted by JNK itself. Furthermore, there are other gene mutations in ovarian tumors such as KRAS,

BRAF, and PTEN, which induce the activation of the JNK signaling pathway to promote growth of cancer cells [62]. In addition, the mutation of RAS can also contribute to the growth of ovarian cancer. RAS family genes including HRAS, KRAS, NRAS are the most common genes to undergo mutations in tumors. Ras^{v12} mutations interact with JNK-Drosophila TGF-β-activated kinase 1(dTAK1) signaling mediating the growth of tumors, contributing to their proliferation out of control behavior [73]. This might be another reason for inducing the rapid growth of ovarian cells but further experiments need to be done to

confirm this. As an important target molecule downstream of JNK, the expression level of c-jun in ovarian cancer tissues is high and is correlated with malignancy [34]. Therefore, inhibiting the signaling pathway of JNK1/c-jun should result in the ability of cellular proliferation and invasion to be significantly decreased [74, 75]. Tumor necrosis factor- α -mediated apoptosis can induce transient activation of JNK and regulate NF- κ B to protect cells from apoptosis [68]. Furthermore, it is positively correlated with alpha 1,2-fucosyltransferase 1/2 (FUT1/2) as well as Lewis y, a sugar of type II. The combination of c-jun and the promoter of FUT1 induce the proliferation of ovarian cancer cells and the complexity of Lewis y/ FUT1 might be regulated by TGF- β 1 [76, 77]. It has been pointed out that c-Fos can enhance the proliferation of ovarian cancer cells induced by TGF- β 1/c-jun [77]. Knocking out c-Fos can down-regulate the expression of cyclin D1 and CD44 as well as inhibit the proliferation and invasion of ovarian cancer cells [77, 78]. Aging human peritoneal mesothelial cells may promote the proliferation and migration of ovarian cancer cells through AP-1/c-jun, IL-6, TGF- β as as well as the phenotypes secreted by tumors, such as angiogenic agents including vascular growth factor and CXCL1 [79].

Moreover, there is also evidence to suggest that the JNK signaling pathway has the function of promoting cancer cell survival in the regulation of cancer cell autophagy and growth [68]. Autophagy can not only induce tumor cell apoptosis as we discussed above, but also induce tumorigenesis and development via the JNK signaling pathway [68, 80]. Many researches have reported that autophagy helps cell survival and proliferation, and in many contexts such as ER stress, chemotherapy, and other diseases can also do so via activating the JNK signal pathway when stimulated for a short time [68, 81]. Different JNK subtypes may have different functions. ER stress can instantaneously activate JNK2 to up-regulate BIP, inhibit cell apoptosis and prevent cell death caused by ER stress, and down-regulate the expression of CHOP, as for transcription factors of cell death and cell apoptosis [68]. Bcl-2, downstream molecular of JNK signaling pathway during the autophagy. Also, the JNK signaling pathway, activated by IRE1 α , may regulate Bcl-2 phosphorylation of Ser70 via JNK1 not JNK2, thus promoting cell survival [40, 82]. Inhibiting the activity of JNK led to the impair of cancer cell development [83]. TP53 has a negative regulation of Bcl-2, inducing the alteration of the pro-apoptosis Bax/Bak, which initiates cell apoptosis [84]. Note that, Ovarian cancer, especially high level serous ovarian cancer, has a high frequency mutation of TP53 [71]. Inhibiting the activation of Bcl-2 contributes to cell apoptosis, which suggests that the JNK1-Bcl-2 signaling pathway is crucial for the autophagy helping cell survival. There is, however, another explanation that JNK mediates the separation of

Beclin1 and Bcl-2/Bcl-XL combinations and induces autophagy. Instantaneous activation of JNK1 can also induce cell survival, but long-term activation can lead to cell apoptosis [85]. When cells are in a hungry state, it is JNK1 rather than JNK2 or JNK3 that mediates the phosphorylation of Bcl-2 so that Beclin1 dissociates from the combinations and stimulates autophagy [40]. Although autophagy-mediating evading apoptosis can help cancer cells maintain a proliferating cell base through intracellular regulation mechanism. Self-regulation mechanism of autophagy may be one of the important reasons leading to drug resistance and recurrence of cancer.

To sum up, the discussion shows that autophagy plays a positive role in the regulation of tumor cell survival via JNK signaling, and gene mutation also promotes tumor growth via activating the JNK pathway. JNK1 induces phosphorylation of Bcl-2, dependent on the length of stimuli, which promotes cells survival. Further research is, therefore, essential to clearly explain the mechanism involved in JNK signaling preventing cell death. Although there are a number of studies in other cancer research areas concerning the promotion of cell survival and growth via the JNK pathway, there are very few in the realm of ovarian cancer and those few only show that inhibiting the JNK pathway impairs the growth of ovarian cancer without demonstrating the underlying mechanism. A potential mechanism, as discussed, for the function in JNK signaling pathway-mediated tumor cell survival and growth is displayed in Fig. 2.

The role of JNK pathway in chemotherapy resistance

The chemotherapy resistance development of cancer cells is one of the important reasons for impairing the effect of drugs, and may be due to an abnormal activation pathway. Such a resistance phenomenon of tumor cells generally occurs after the initial chemotherapy is given, and in the later stage, drug resistance, aggravation, followed by recurrence may happen in a process similar to Darwin's natural selection rules, the survival of the fittest. As the drug time increases, the resistance of tumor cells will gradually increase. Chemotherapy kills most ovarian cancer cells, but some cancer stem cells with unique hereditary characteristics survive and represent a hidden danger for future recurrence and drug resistance [86]. In addition, the body's immune system is also a very important accomplice for the recurrence of ovarian cancer, in that through the use of immune cell recognition functions by tumor cells, they can modify their own immunogenicity, namely cancer immune editing [87, 88]. For an instance, cisplatin, a first line chemotherapy drug for the treatment of advanced ovarian tumors, induces the rate of cancer cell death up to more than 50% but the subsequent treatment effect is decreased owing to drug

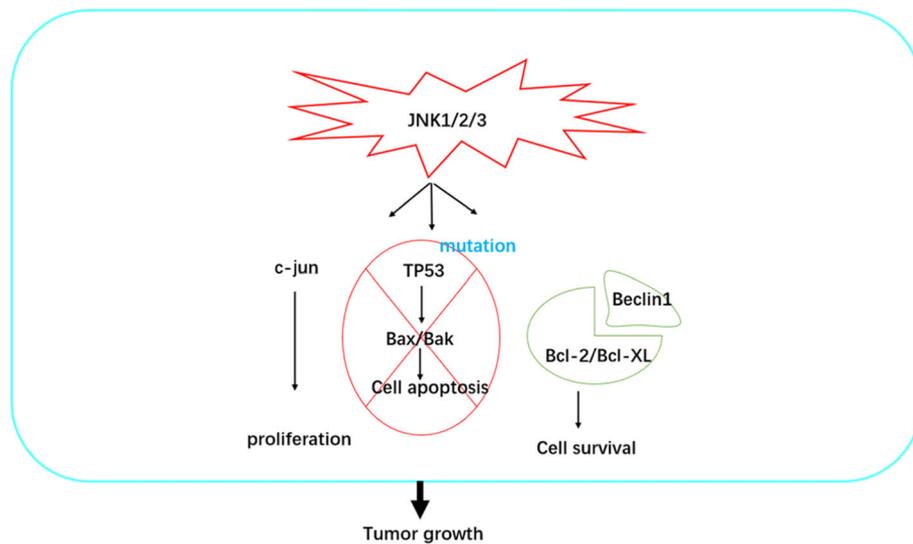


Fig. 2 A schematic diagram of the JNK signaling pathway promoting tumor growth in ovarian cancer. C-Jun promotes the proliferation of tumors; JNK-mediated p53 pathway can induce apoptosis, but the mutation of p53 in ovarian cancer can prevent the death of tumors; in addition, JNK signaling pathway

resistance or recurrence after a long time of chemotherapy exposure [89].

Therefore, in order to look for potential therapeutic targets of chemotherapy resistance to improve the effect of treatment, many researches focused on studying the mechanism of chemotherapy drug resistance. The activation of the JNK signaling pathway plays an important role in promoting drug resistance in cancer [90, 91]. Furthermore, there are also many researches reporting that c-jun is overexpressed in cisplatin resistance in ovarian cancer tissue [34]. The negative expression of c-jun mutation can enhance the sensitivity of the human ovarian cancer cells Caov-3 and A2780 to cisplatin [92]. JNK activity is higher in cisplatin or paclitaxel-resistant human ovarian cancer cell lines and has the positive correlation with drug resistance. In addition, the high expression of active JNK is reported to have a close relationship with stage III and stage IV compared with stage I and stage II, which also was negatively correlated with the survival rate of patients with ovarian cancer [92]. Activation of JNK may play a key role in the DNA repair mechanism due to cisplatin therapy. When cisplatin and SP600125 were combined to treat ovarian cancer cells, the anti-cancer effect was improved compared with cisplatin alone. Compared with the combination of paclitaxel and SP600125, the anti-cancer effect of paclitaxel is weakened, because the death signal caused by paclitaxel is related to the activity of JNK. However, pretreatment of ovarian cancer cells with SP600125 can improve the anti-cancer effects of cisplatin and paclitaxel [92]. Platinum-based drugs increase the level of ROS via the mitochondrial apoptosis

signaling pathway. ROS, however, damages the structure of DNA, Protein and other molecular integrity. Note that, sirtuin6, phosphorylated at serine 10 by the JNK pathway, modifies Kap1 so as to promote the silence of L1 retrotransposons [93], and SIRT6 also mediating single adenosine diphosphate ribosylation of PARP1 increases the activity of PARP1 poly adenosine diphosphate ribosylation and promotes DNA double strand break repair [93]. It may also be related to JNK-induced autophagy or gene mutation of tumors themselves helping cells survive. After all, JNK can induce both autophagy and apoptosis by regulating the interaction between Beclin1-Bcl2 complex and Bcl2-Bax [94, 95]. Activation of JNK leads to the destruction of the Bax-Bcl2 complex, which increases the expression of Bax promoting cell apoptosis [96]. JNK-induced autophagy depends on the dissociation of Beclin1, which comes from the Beclin1-Bcl2 complex by phosphorylation of Bcl2. The production of ROS results in mitochondrial damage. There is a small positive regulatory loop between activating apoptosis; ROS, JNK and p53, that is, ROS can activate the JNK signaling pathway. Separation of the p53-MDM2 complex can lead to ROS accumulation, while JNK can activate the downstream gene p53 [95]. The ROS/JNK signaling pathway can promote rapid cell apoptosis through p53; however, mutation of p53 in some ovarian cancer tissue may avoid ROS/JNK/p53 signaling pathway mediated rapid apoptosis of cells.

Micro RNAs (miRNA), having the length of about 21 nucleotides, do not participate in coding protein directly; these are also be called non-coding RNAs. MiRNAs regulate the expression of gene transcription and also have an association with chemotherapy drug resistance as well as

the progression and oncogenesis of cancer. MiR-21, over-expressed highly in many cancers such as breast cancer and ovarian cancer, induces the resistance of cisplatin or paclitaxel [75, 97]. The promoter region of pri-miR-21 interacts with p-c-jun specifically and c-jun is activated by JNK1 not JNK2 or JNK3 in the chemotherapy resistance of ovarian tumor cells [75]. MiR-21 inhibits the expression of programmed cell death 4 (PDCD4), which is directly related to poor prognosis of ovarian tumor patients [98]. Besides, miR-21 also regulates the expression of hypoxia inducible factor-1 α influencing tumor cell metabolism [99]. Fra-1, a member of the Fos family, form dimer AP-1 with the Jun family. AP-1 regulates miR-134 so as to augment the function of the JNK signaling pathway mediating chemotherapy insensitivity of ovarian tumor cells [100]. However, not all miRNAs promote the development of tumor chemotherapy resistance. For example, miR-139-5p can interact with c-jun by binding site 3' UTR of mRNA, which interferes with the combination of c-jun and ATF2 induced by cisplatin and also inhibits the expression of Bcl-xl in cisplatin resistance cells [101].

These researches suggest that the JNK signaling pathway plays an essential role in chemotherapy resistance of ovarian cancer cells, although its functions are contradictory in tumors. Autophagy mediated by the JNK signaling pathway may provide resistance to chemotherapeutic drugs, leading to drug resistance in the later stage of relapse. Mutations in the P53 gene may inhibit apoptosis induced by ROS/JNK/p53, although the JNK pathway may induce apoptosis in a caspase-dependent manner. Studies have shown that the staggered use of JNK inhibitors and

the chemotherapeutic drugs platinum or paclitaxel in turn can increase chemotherapeutic sensitivity without increasing side effects. The mechanism of JNK-mediated chemotherapeutic drug resistance discussed above is shown in Fig. 3. It is clear that JNK-mediated drug resistance in ovarian cancer chemotherapy is a difficult problem that needs to be solved as soon as possible.

Conclusion

It is clear that understanding dual role of the JNK signaling pathway, that can both promote the development of tumorigenesis and also inhibit the progress of tumors, the so called seesaw role, is very important in the study of cancer. The immune escape mechanism of tumors may also be involved in JNK-mediated tumor cell survival, such as interleukin and tumor necrosis factor. Although many studies have provided detailed evidence for JNK to promote the survival of cancer cells and inhibit the development of cancer, many problems remain unsolved, including the following: (1) JNK signaling pathway mediates death in the early stage and induces drug resistance in the later stage. Besides the reason of activation time and phosphorylation SIRT6 so as to repair DNA breaks, how it specifically affects JNK signaling pathway has not been confirmed. And vivo experiments and clinical studies need to be improved. (2) JNK1 can promote cell proliferation, and autophagy induced by external stimulation of the JNK signaling pathway and can also help cancer cells avoid apoptosis induced by drugs. But under what stress conditions these external stimuli and chemotherapeutics cause

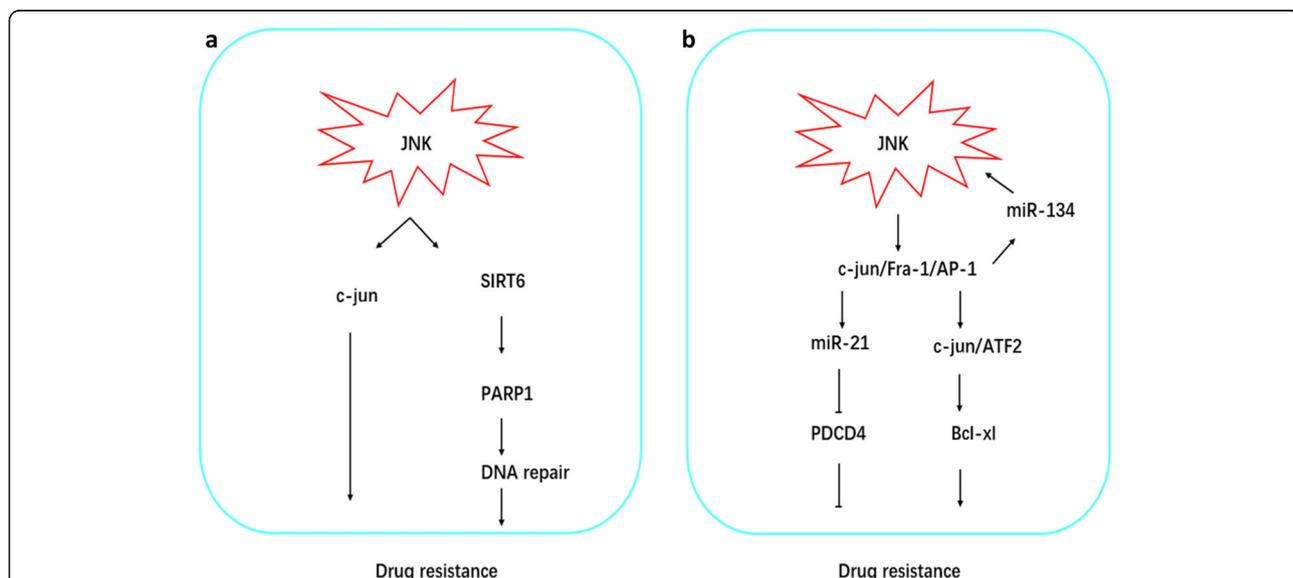


Fig. 3 JNK signaling pathway mediated drug resistance. **a** NK signaling pathway mediated by the JNK signaling pathway in ovarian cancer can repair DNA damage through SIRT6/PARP1 or reduce the sensitivity of ovarian cancer tissues to chemotherapy by c-Jun. **b** Drug resistance mediated by JNK signaling pathway also involves microRNA. MiR-21 inhibits the expression of PDCD4 and promotes resistance, or the interaction between miR-134 and JNK signaling pathway promotes the activation of c-jun/ATF2 and promotes drug resistance

autophagic apoptosis rather than autophagy. (3) At present, the evidence of many oncology experiments related to the JNK signaling pathway comes from cell experiments, especially on ovarian cancer cells. The study of mouse models and clinical trials will help to improve our understanding of the function of the JNK signaling pathway in tumors. Because the occurrence and development of cancer is affected by the overall internal environment of the body, not a single factor.

The JNK signaling pathway regulates both cell death and survival. It is, therefore, important to study the role of the JNK signaling pathway in each of these conflicting situations and the mechanisms involved in “see-sawing” between each other. In the future, it is necessary to determine the specific relationship between JNK structure and function. In view of this though, we firmly believe that in the near future, defining the specific mechanism of JNK in the process of tumorigenesis and development, and targeting the JNK signaling pathway in cancer treatment-related drugs will provide tremendous help to spearhead a breakthrough in anti-cancer treatment.

Abbreviations

ASK1: Apoptosis signal regulation kinase 1; ER stress: Endoplasmic reticulum stress; JNK: Jun N-terminal kinase; MAPK: Mitogen-activated protein kinases; OC: Ovarian carcinoma; ROS: reactive oxygen species

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Authors' contributions

YD proposed the idea and was a major contributor in writing the manuscript. XJ, HX and JH collected related-article. SX revised the article. All authors read and approved the final manuscript.

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Competing interests

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Author details

¹Department of Gynecology and Obstetrics, the third Xiangya Hospital, the Central South University, Changsha 410013, Hunan, China. ²Cancer Research Institute, the Central South University, Changsha 410011, Hunan, China.

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References

- Reid BM, Permuth JB, Sellers TA. Epidemiology of ovarian cancer: a review. *Cancer Biol Med.* 2017;14(1):9–32.
- Ferlay J, Soerjomataram I, Dikshit R, Eser S, Mathers C, Rebelo M, Parkin DM, Forman D, Bray F. Cancer incidence and mortality worldwide: sources, methods and major patterns in GLOBOCAN 2012. *Int J Cancer.* 2015;136(5):E359–86.
- Torre LA, Trabert B, DeSantis CE, Miller KD, Samimi G, Runowicz CD, Gaudet MM, Jemal A, Siegel RL. Ovarian cancer statistics, 2018. *CA Cancer J Clin.* 2018;68(4):284–96.
- Chen W, Zheng R, Baade PD, Zhang S, Zeng H, Bray F, Jemal A, Yu XQ, He J. Cancer statistics in China, 2015. *CA Cancer J Clin.* 2016;66(2):115–32.
- Burges A, Schmalfeldt B. Ovarian cancer: diagnosis and treatment. *Dtsch Arztebl Int.* 2011;108(38):635–41.
- Hirst J, Crow J, Godwin A. Ovarian cancer genetics: subtypes and risk factors; 2018.
- Oberaigner W, Minicozzi P, Bielska-Lasota M, Allemanni C, de Angelis R, Mangone L, Sant M, Eurocare Working G. Survival for ovarian cancer in Europe: the across-country variation did not shrink in the past decade. *Acta Oncol.* 2012;51(4):441–53.
- Maas HA, Kruitwagen RF, Lemmens VE, Goey SH, Janssen-Heijnen ML. The influence of age and co-morbidity on treatment and prognosis of ovarian cancer: a population-based study. *Gynecol Oncol.* 2005;97(1):104–9.
- Schuurman MS, Kruitwagen R, Portielje JEA, Roes EM, Lemmens V, van der Aa MA. Treatment and outcome of elderly patients with advanced stage ovarian cancer: a nationwide analysis. *Gynecol Oncol.* 2018;149(2):270–4.
- Jorgensen TL, Teiblum S, Paludan M, Poulsen LO, Jorgensen AY, Bruun KH, Hallas J, Herrstedt J. Significance of age and comorbidity on treatment modality, treatment adherence, and prognosis in elderly ovarian cancer patients. *Gynecol Oncol.* 2012;127(2):367–74.
- Vitale SG, Capriiglione S, Zito G, Lopez S, Gulino FA, Di Guardo F, Vitagliano A, Noventa M, La Rosa VL, Sapia F, et al. Management of endometrial, ovarian and cervical cancer in the elderly: current approach to a challenging condition. *Arch Gynecol Obstet.* 2019;299(2):299–315.
- Bailey MH, Tokheim C, Porta-Pardo E, Sengupta S, Bertrand D, Weerasinghe A, Colaprico A, Wendl MC, Kim J, Reardon B, et al. Comprehensive characterization of cancer driver genes and mutations. *Cell.* 2018;173(2):371–85 e318.
- Vogelstein B, Papadopoulos N, Velculescu VE, Zhou S, Diaz LA Jr, Kinzler KW. Cancer genome landscapes. *Science.* 2013;339(6127):1546–58.
- Burotto M, Chiou VL, Lee JM, Kohn EC. The MAPK pathway across different malignancies: a new perspective. *Cancer.* 2014;120(22):3446–56.
- Johnson GL, Nakamura K. The c-jun kinase/stress-activated pathway: regulation, function and role in human disease. *Biochim Biophys Acta.* 2007;1773(8):1341–8.
- Jo H, Ahn HJ, Lee JH, Min CK. Roles of JNK and P53 in taxol-induced apoptotic signaling in SKOV3 human ovarian cancer cells. *Elyns J Cancer Res.* 2016;01(01). <https://doi.org/10.19104/ejcr.2015.101>
- Seino M, Okada M, Shibuya K, Seino S, Suzuki S, Ohta T, Kurachi H, Kitanaka C. Requirement of JNK signaling for self-renewal and tumor-initiating capacity of ovarian cancer stem cells. *Anticancer Res.* 2014;34(9):4723–31.
- Johnson GL, Lapadat R. Mitogen-activated protein kinase pathways mediated by ERK, JNK, and p38 protein kinases. *Science.* 2002;298(5600):1911–2.
- Davis RJ. Signal transduction by the JNK group of MAP kinases. *Cell.* 2000;103(2):239–52.
- Wang X, Destrument A, Tournier C. Physiological roles of MKK4 and MKK7: insights from animal models. *Biochim Biophys Acta.* 2007;1773(8):1349–57.
- Hayakawa R, Hayakawa T, Takeda K, Ichijo H. Therapeutic targets in the ASK1-dependent stress signaling pathways. *Proc Jpn Acad Ser B.* 2012;88(8):434–53.
- Ma FY, Tesch GH, Nikolic-Paterson DJ. ASK1/p38 signaling in renal tubular epithelial cells promotes renal fibrosis in the mouse obstructed kidney. *Am J Physiol Renal Physiol.* 2014;307(11):F1263–73.
- Maurya SK, Mishra J, Tripathi VK, Sharma R, Siddiqui MH. Cypermethrin induces astrocyte damage: role of aberrant Ca²⁺, ROS, JNK, P38, matrix metalloproteinase 2 and migration related reelin protein. *Pestic Biochem Physiol.* 2014;111:51–9.
- Seki E, Brenner DA, Karin M. A liver full of JNK: signaling in regulation of cell function and disease pathogenesis, and clinical approaches. *Gastroenterology.* 2012;143(2):307–20.
- Corazza N, Jakob S, Schaer C, Frese S, Keogh A, Stroka D, Kassahn D, Torgler R, Mueller C, Schneider P, et al. TRAIL receptor-mediated JNK activation and Bim phosphorylation critically regulate Fas-mediated liver damage and lethality. *J Clin Invest.* 2006;116(9):2493–9.
- Kim HS, Asmis R. Mitogen-activated protein kinase phosphatase 1 (MKP-1) in macrophage biology and cardiovascular disease. A redox-regulated master controller of monocyte function and macrophage phenotype. *Free Radic Biol Med.* 2017;109:75–83.

27. Goldman EH, Chen L, Fu H. Activation of apoptosis signal-regulating kinase 1 by reactive oxygen species through dephosphorylation at serine 967 and 14-3-3 dissociation. *J Biol Chem*. 2004;279(11):10442–9.
28. Li L, Feng Z, Porter AG. JNK-dependent phosphorylation of c-Jun on serine 63 mediates nitric oxide-induced apoptosis of neuroblastoma cells. *J Biol Chem*. 2004;279(6):4058–65.
29. Maeda S, Karin M. Oncogene at last—c-Jun promotes liver cancer in mice. *Cancer Cell*. 2003;3(2):102–4.
30. Tournier C. The 2 faces of JNK signaling in cancer. *Genes Cancer*. 2013;4(9–10):397–400.
31. Leppa S, Saffrich R, Ansorge W, Bohmann D. Differential regulation of c-Jun by ERK and JNK during PC12 cell differentiation. *EMBO J*. 1998;17(15):4404–13.
32. Win S, Than TA, Zhang J, Oo C, Min RWM, Kaplowitz N. New insights into the role and mechanism of c-Jun-N-terminal kinase signaling in the pathobiology of liver diseases. *Hepatology*. 2018;67(5):2013–24.
33. Win S, Than TA, Fernandez-Checa JC, Kaplowitz N. JNK interaction with Sab mediates ER stress induced inhibition of mitochondrial respiration and cell death. *Cell Death Dis*. 2014;5:e989.
34. Lee CM, Lo H-W, Shao R-P, Wang S-C, Xia W, Gershenson DM, Hung M-C. Selective activation of ceruloplasmin promoter in ovarian tumors. *Cancer Res*. 2004;64(5):1788–93.
35. Huang L, Jin Y, Feng S, Zou Y, Xu S, Qiu S, Li L, Zheng J. Role of Wnt/beta-catenin, Wnt/c-Jun N-terminal kinase and Wnt/Ca (2+) pathways in cisplatin-induced chemoresistance in ovarian cancer. *Exp Ther Med*. 2016;12(6):3851–8.
36. Dhanasekaran DN, Reddy EP. JNK signaling in apoptosis. *Oncogene*. 2008;27(48):6245–51.
37. Achkar IW, Abdulrahman N, Al-Sulaiti H, Joseph JM, Uddin S, Mraiche F. Cisplatin based therapy: the role of the mitogen activated protein kinase signaling pathway. *J Transl Med*. 2018;16(1). <https://doi.org/10.1186/s12967-018-1471-1>
38. Hanahan D, Weinberg RA. Hallmarks of cancer: the next generation. *Cell*. 2011;144(5):646–74.
39. Glick D, Barth S, Macleod KF. Autophagy: cellular and molecular mechanisms. *J Pathol*. 2010;221(1):3–12.
40. Wei Y, Pattinre S, Sinha S, Bassik M, Levine B. JNK1-mediated phosphorylation of Bcl-2 regulates starvation-induced autophagy. *Mol Cell*. 2008;30(6):678–88.
41. Lu Z, Miao Y, Muhammad I, Tian E, Hu W, Wang J, Wang B, Li R, Li J. Colistin-induced autophagy and apoptosis involves the JNK-Bcl2-Bax signaling pathway and JNK-p53-ROS positive feedback loop in PC-12 cells. *Chem Biol Interact*. 2017;277:62–73.
42. Bhattacharjee A, Hasanain M, Kathuria M, Singh A, Datta D, Sarkar J, Mitra K. Ormeloxifene-induced unfolded protein response contributes to autophagy-associated apoptosis via disruption of Akt/mTOR and activation of JNK. *Sci Rep*. 2018;8(1):2303.
43. Wang Y, Singh R, Xiang Y, Czaja MJ. Macroautophagy and chaperone-mediated autophagy are required for hepatocyte resistance to oxidant stress. *Hepatology*. 2010;52(1):266–77.
44. Xu L, Zhang X, Li Y, Lu S, Lu S, Li J, Wang Y, Tian X, Wei JJ, Shao C, et al. Neferine induces autophagy of human ovarian cancer cells via p38 MAPK/JNK activation. *Tumour Biol*. 2016;37(7):8721–9.
45. Zhang K, Kaufman RJ. Signaling the unfolded protein response from the endoplasmic reticulum. *J Biol Chem*. 2004;279(25):25935–8.
46. Kim R, Emi M, Tanabe K, Murakami S. Role of the unfolded protein response in cell death. *Apoptosis*. 2006;11(1):5–13.
47. Luo B, Lee AS. The critical roles of endoplasmic reticulum chaperones and unfolded protein response in tumorigenesis and anticancer therapies. *Oncogene*. 2013;32(7):805–18.
48. Zheng GF, Cai Z, Meng XK, Zhang Y, Zhu W, Pang XY, Dou L. Unfolded protein response mediated JNK/AP-1 signal transduction, a target for ovarian cancer treatment. *Int J Clin Exp Pathol*. 2015;8(6):6505–11.
49. Yoneda T, Imaizumi K, Oono K, Yui D, Gomi F, Katayama T, Tohyama M. Activation of caspase-12, an endoplasmic reticulum (ER) resident caspase, through tumor necrosis factor receptor-associated factor 2-dependent mechanism in response to the ER stress. *J Biol Chem*. 2001;276(17):13935–40.
50. Tong Q, Wu L, Gao Q, Ou Z, Zhu D, Zhang Y. PPARbeta/delta agonist provides neuroprotection by suppression of IRE1alpha-caspase-12-mediated endoplasmic reticulum stress pathway in the rotenone rat model of Parkinson's disease. *Mol Neurobiol*. 2016;53(6):3822–31.
51. Ma L, Wei J, Wan J, Wang W, Wang L, Yuan Y, Yang Z, Liu X, Ming L. Low glucose and metformin-induced apoptosis of human ovarian cancer cells is connected to ASK1 via mitochondrial and endoplasmic reticulum stress-associated pathways. *J Exp Clin Cancer Res*. 2019;38(1):77.
52. Amos LA, Ma FY, Tesch GH, Liles JT, Breckenridge DG, Nikolic-Paterson DJ, Han Y. ASK1 inhibitor treatment suppresses p38/JNK signalling with reduced kidney inflammation and fibrosis in rat crescentic glomerulonephritis. *J Cell Mol Med*. 2018;22(9):4522–33.
53. Ganapathy-Kanniappan S, Geschwind JFH. Tumor glycolysis as a target for cancer therapy: progress and prospects. *Mol Cancer*. 2013;12. <http://www.molecular-cancer.com/content/12/1/152>
54. Kajihara N, Kukidome D, Sada K, Motoshima H, Furukawa N, Matsumura T, Nishikawa T, Araki E. Low glucose induces mitochondrial reactive oxygen species via fatty acid oxidation in bovine aortic endothelial cells. *J Diabetes Investig*. 2017;8(6):750–61.
55. Choudhury S, Bae S, Ke Q, Lee JY, Kim J, Kang PM. Mitochondria to nucleus translocation of AIF in mice lacking Hsp70 during ischemia/reperfusion. *Basic Res Cardiol*. 2011;106(3):397–407.
56. Sharifi S, Barar J, Hejazi MS, Samadi N. Doxorubicin changes Bax /Bcl-xL ratio, caspase-8 and 9 in breast cancer cells. *Adv Pharm Bull*. 2015;5(3):351–9.
57. Hata AN, Engelman JA, Faber AC. The BCL2 family: key mediators of the apoptotic response to targeted anticancer therapeutics. *Cancer Discov*. 2015;5(5):475–87.
58. Shen K, Xie J, Wang H, Zhang H, Yu M, Lu F, Tan H, Xu H. Cambogin induces caspase-independent apoptosis through the ROS/JNK pathway and epigenetic regulation in breast cancer cells. *Mol Cancer Ther*. 2015;14(7):1738–49.
59. Chen YJ, Liu WH, Kao PH, Wang JJ, Chang LS. Involvement of p38 MAPK- and JNK-modulated expression of Bcl-2 and Bax in Naja nigricollis CMS-9-induced apoptosis of human leukemia K562 cells. *Toxicol*. 2010;55(7):1306–16.
60. Cui ZY, Park SJ, Jo E, Hwang JH, Lee KB, Kim SW, Kim DJ, Joo JC, Hong SH, Lee MG, et al. Cordycepin induces apoptosis of human ovarian cancer cells by inhibiting CCL5-mediated Akt/NF-kappaB signaling pathway. *Cell Death Discov*. 2018;4:62.
61. Chen TC, Yu MC, Chien CC, Wu MS, Lee YC, Chen YC. Nilotinib reduced the viability of human ovarian cancer cells via mitochondria-dependent apoptosis, independent of JNK activation. *Toxicol in Vitro*. 2016;31:1–11.
62. Vivas-Mejia P, Benito JM, Fernandez A, Han HD, Mangala L, Rodriguez-Aguayo C, Chavez-Reyes A, Lin YG, Carey MS, Nick AM, et al. c-Jun-NH2-kinase-1 inhibition leads to antitumor activity in ovarian cancer. *Clin Cancer Res*. 2010;16(1):184–94.
63. du Bois A, Kristensen G, Ray-Coquard I, Reuss A, Pignata S, Colombo N, Denison U, Vergote I, del Campo JM, Ottavanger P, et al. Standard first-line chemotherapy with or without nintedanib for advanced ovarian cancer (AGO-OVAR 12): a randomised, double-blind, placebo-controlled phase 3 trial. *Lancet Oncol*. 2016;17(1):78–89.
64. Barra F, Lagana AS, Ghezzi F, Casarin J, Ferrero S. Nintedanib for advanced epithelial ovarian cancer: a change of perspective? Summary of evidence from a systematic review. *Gynecol Obstet Investig*. 2019;84(2):107–17.
65. Kamio K, Usuki J, Azuma A, Matsuda K, Ishii T, Inomata M, Hayashi H, Kokuho N, Fujita K, Saito Y, et al. Nintedanib modulates surfactant protein-D expression in A549 human lung epithelial cells via the c-Jun-N-terminal kinase-activator protein-1 pathway. *Pulm Pharmacol Ther*. 2015;32:29–36.
66. Liu J, Wang L, Zhang Y, Li S, Sun F, Wang G, Yang T, Wei D, Guo L, Xiao H. Induction of entosis in prostate cancer cells by nintedanib and its therapeutic implications. *Oncol Lett*. 2019;17(3):3151–62.
67. Kuan CY, Yang DD, Samanta Roy DR, Davis RJ, Rakic P, Flavell RA. The Jnk1 and Jnk2 protein kinases are required for regional specific apoptosis during early brain development. *Neuron*. 1999;22(4):667–76.
68. Raciti M, Lotti LV, Valia S, Pulcinelli FM, Di Renzo L. JNK2 is activated during ER stress and promotes cell survival. *Cell Death Dis*. 2012;3:e429.
69. Hanahan D, Weinberg RA. The hallmarks of cancer. *Cell*. 2000;100(1):57–70.
70. Bowtell DD, Bohm S, Ahmed AA, Aspuria PJ, Bast RC Jr, Beral V, Berek JS, Birrer MJ, Blagden S, Bookman MA, et al. Rethinking ovarian cancer II: reducing mortality from high-grade serous ovarian cancer. *Nat Rev Cancer*. 2015;15(11):668–79.
71. Zhang Y, Cao L, Nguyen D, Lu H. TP53 mutations in epithelial ovarian cancer. *Transl Cancer Res*. 2016;5(6):650–63.
72. Ahmed AA, Etemadmoghadam D, Temple J, Lynch AG, Riad M, Sharma R, Stewart C, Feraday S, Caldas C, Defazio A, et al. Driver mutations in TP53 are ubiquitous in high grade serous carcinoma of the ovary. *J Pathol*. 2010;221(1):49–56.

73. Ma X, Lu JY, Dong Y, Li D, Malagon JN, Xu T. PP6 disruption synergizes with oncogenic Ras to promote JNK-dependent tumor growth and invasion. *Cell Rep*. 2017;19(13):2657–64.
74. Lim V, Zhu H, Diao S, Hu L, Hu J. PKP3 interactions with MAPK-JNK-ERK1/2-mTOR pathway regulates autophagy and invasion in ovarian cancer. *Biochem Biophys Res Commun*. 2019;508(2):646–53.
75. Echevarria-Vargas IM, Valiyeva F, Vivas-Mejia PE. Upregulation of miR-21 in cisplatin resistant ovarian cancer via JNK-1/c-Jun pathway. *PLoS One*. 2014; 9(5):e97094.
76. Gao N, Liu J, Liu D, Hao Y, Yan L, Ma Y, Zhuang H, Hu Z, Gao J, Yang Z, et al. c-Jun transcriptionally regulates alpha 1, 2-fucosyltransferase 1 (FUT1) in ovarian cancer. *Biochimie*. 2014;107 Pt B:286–92.
77. Hao Y, Zhu L, Yan L, Liu J, Liu D, Gao N, Tan M, Gao S, Lin B. c-Fos mediates alpha1, 2-fucosyltransferase 1 and Lewis y expression in response to TGF-beta1 in ovarian cancer. *Oncol Rep*. 2017;38(6):3355–66.
78. Dong C, Ye DX, Zhang WB, Pan HY, Zhang ZY, Zhang L. Overexpression of c-fos promotes cell invasion and migration via CD44 pathway in oral squamous cell carcinoma. *J Oral Pathol Med*. 2015;44(5):353–60.
79. Mikula-Pietrasik J, Sosinska P, Naumowicz E, Maksin K, Piotrowska H, Wozniak A, Szperek D, Ksiazek K. Senescent peritoneal mesothelium induces a pro-angiogenic phenotype in ovarian cancer cells in vitro and in a mouse xenograft model in vivo. *Clin Exp Metastasis*. 2016;33(1):15–27.
80. Wu Q, Wu W, Fu B, Shi L, Wang X, Kuca K. JNK signaling in cancer cell survival. *Med Res Rev*. 2019. <https://doi.org/10.1002/med.21574>
81. Papa S, Zazzeroni F, Pham CG, Bubicic C, Franzoso G. Linking JNK signaling to NF-kappaB: a key to survival. *J Cell Sci*. 2004;117(Pt 22):5197–208.
82. Deng X, Xiao L, Lang W, Gao F, Ruvolo P, May WS Jr. Novel role for JNK as a stress-activated Bcl2 kinase. *J Biol Chem*. 2001;276(26):23681–8.
83. Yuan D, Huang S, Berger E, Liu L, Gross N, Heinzmann F, Ringelhan M, Connor TO, Stadler M, Meister M, et al. Kupffer cell-derived Tnf triggers cholangiocellular tumorigenesis through JNK due to chronic mitochondrial dysfunction and ROS. *Cancer Cell*. 2017;31(6):771–89 e776.
84. Campbell KJ, Tait SWG. Targeting BCL-2 regulated apoptosis in cancer. *Open Biol*. 2018;8(5). <https://doi.org/10.1098/rsob.180002>
85. Ham Y-M, Chun K-H, Choi J-S, Kim D-H, Lee S-K. SEK1-dependent JNK1 activation prolongs cell survival during G-Rh2-induced apoptosis. *Biochem Biophys Res Commun*. 2003;304(2):358–64.
86. Laganà AS, Colonese F, Colonese E, Sofo V, Salmeri FM, Granese R, Chiofalo B, Ciancimino L, Triolo O. Cytogenetic analysis of epithelial ovarian cancer's stem cells: an overview on new diagnostic and therapeutic perspectives. *Eur J Gynaecol Oncol*. 2015;36(5):495–505.
87. Schreiber RD, Old LJ, Smyth MJ. Cancer immunoediting: integrating immunity's roles in cancer suppression and promotion. *Science*. 2011; 331(6024):1565–70.
88. Laganà AS, Sofo V, Vitale SG, Triolo O. Epithelial ovarian cancer inherent resistance: may the pleiotropic interaction between reduced immunosurveillance and drug-resistant cells play a key role? *Gynecol Oncol Rep*. 2016;18:57–8.
89. Dasari S, Tchounwou PB. Cisplatin in cancer therapy: molecular mechanisms of action. *Eur J Pharmacol*. 2014;740:364–78.
90. Sui X, Kong N, Wang X, Fang Y, Hu X, Xu Y, Chen W, Wang K, Li D, Jin W, et al. JNK confers 5-fluorouracil resistance in p53-deficient and mutant p53-expressing colon cancer cells by inducing survival autophagy. *Sci Rep*. 2014;4:4694.
91. Ferrao PT. Taking out the JNK: a window of opportunity to improve cancer therapy. *Mol Cell Oncol*. 2016;3(3):e1128515.
92. Seino M, Okada M, Sakaki H, Takeda H, Watarai H, Suzuki S, Seino S, Kuramoto K, Ohta T, Nagase S, et al. Time-staggered inhibition of JNK effectively sensitizes chemoresistant ovarian cancer cells to cisplatin and paclitaxel. *Oncol Rep*. 2016;35(1):593–601.
93. Van Meter M, Simon M, Tomblin G, May A, Morello TD, Hubbard BP, Bredbenner K, Park R, Sinclair DA, Bohr VA, et al. JNK phosphorylates SIRT6 to stimulate DNA double-strand break repair in response to oxidative stress by recruiting PARP1 to DNA breaks. *Cell Rep*. 2016;16(10):2641–50.
94. Marquez RT, Xu L. Bcl-2: Beclin 1 complex: multiple mechanisms regulating autophagy/apoptosis toggle switch. *Am J Cancer Res*. 2012;2(2):214–21.
95. Decuyper JP, Parys JB, Bultynck G. Regulation of the autophagic bcl-2/beclin 1 interaction. *Cells*. 2012;1(3):284–312.
96. Yang J, Yao S. JNK-Bcl-2/Bcl-xL-Bax/Bak pathway mediates the crosstalk between matrine-induced autophagy and apoptosis via interplay with beclin 1. *Int J Mol Sci*. 2015;16(10):25744–58.
97. Yan LX, Huang XF, Shao Q, Huang MY, Deng L, Wu QL, Zeng YX, Shao JY. MicroRNA miR-21 overexpression in human breast cancer is associated with advanced clinical stage, lymph node metastasis and patient poor prognosis. *RNA*. 2008;14(11):2348–60.
98. Gaur AB, Holbeck SL, Colburn NH, Israel MA. Downregulation of Pdc4 by mir-21 facilitates glioblastoma proliferation in vivo. *Neuro-Oncology*. 2011; 13(6):580–90.
99. Chen S, Chen X, Shan T, Ma J, Lin W, Li W, Kang Y. MiR-21-mediated metabolic alteration of cancer-associated fibroblasts and its effect on pancreatic cancer cell behavior. *Int J Biol Sci*. 2018;14(1):100–10.
100. Wu J, Sun Y, Zhang PY, Qian M, Zhang H, Chen X, Ma D, Xu Y, Chen X, Tang KF. The Fra-1-miR-134-SDS22 feedback loop amplifies ERK/JNK signaling and reduces chemosensitivity in ovarian cancer cells. *Cell Death Dis*. 2016;7(9):e2384.
101. Jiang Y, Jiang J, Jia H, Qiao Z, Zhang J. Recovery of miR-139-5p in ovarian cancer reverses cisplatin resistance by targeting C-Jun. *Cell Physiol Biochem*. 2018;51(1):129–41.

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